



YB
Yeastern Biotech

Product Catalog



YEASTERN BIOTECH

Timesaving
Easy
Accurate



MISSION

We are a Taiwan based molecular biology kits & reagents manufacturer which was founded in 2000 with the goal of providing the intelligent technical services and high quality products in life sciences and in the biomedical industry. We believe that a scientist's greatest help comes from fellow scientists. Therefore, our innovative interdisciplinary research team with highly qualified experts in different fields offers ODM or OEM research-only reagents and kits as well as diagnostic products.

AIM & PROSPECT

The spirits of innovation, integration and humanism in biotechnology are our executive prospects. Based on our possession of excellent research capability and professional technical platforms, we head for long-term aims at developing our own brand of life science and biomedical products.

STRATEGY

We are ISO9001, ISO13485 & GMP certified. Our guideline for product development is to offer time-saving, easy, and accurate (T.E.A.) products to meet our customers' needs. To promote our products and technical services, we strive towards establishing strategic alliance and collaboration with our distributors. We believe we are making impacts on the global biotech market with our innovative products and technology through sustainable and mutual partnership.



ISO 9001 | ISO 13485



Yeastern Biotech Co., Ltd.

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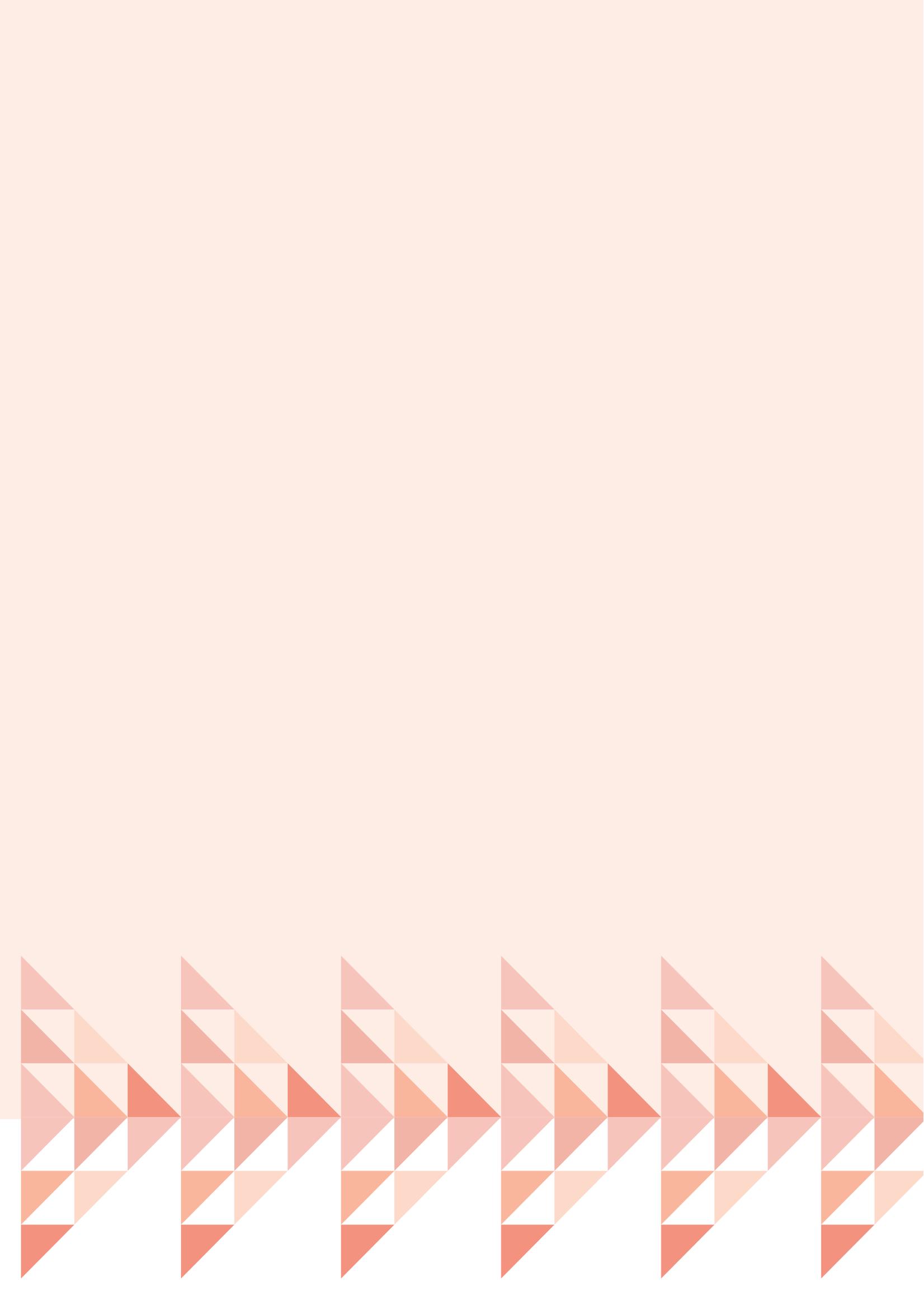
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DNA/RNA Extraction

UniversAll™ Tissue Extraction/PCR Kits



FYU002-5ML/50ML

UniversAll™ Plants Tissue Extraction Buffer

FYU003-20ML

UniversAll™ Animal Tissue Extraction Buffer

FYU009-5ML/20ML

UniversAll™ Robuster Extraction Buffer

Description

The UniversAll™ Tissue PCR Kits contain all the reagents necessary to rapidly extract genomic DNA from a wide range of biological samples prior to amplifying targets of interest by PCR. Genomic DNA is extracted from a small amount of samples simply by incubation in UniversAll™ Extract buffer for 10 minutes at 95 °C. The UniversAll™ Tissue PCR Kits offer a novel one-step UniversAll™ Extract buffer that eliminates the need for freezing cells or tissues with liquid nitrogen, mechanical disruption, organic extraction, column DNA purification, or alcohol precipitation.

Features

- **Simple:** single-step extraction of genomic DNA prior to PCR. No phenol/chloroform.
- **Fast:** cells or tissues to PCR in 10 minutes.
- **Convenient:** includes our superior PCR enzymes and buffers for amplification directly from the extract for your convenience.

Applications

- Gene cloning
- Small or large scale PCR-based genotyping
- Traditional PCR; real-time PCR
- Suitable for rapid forensic and disease detection and diagnosis

Quality Control

Each lot of extraction buffer was tested for its extraction capability of standard animal and plant tissues for PCR and real-time PCR. It also has to pass the stability test before shipping.

Results



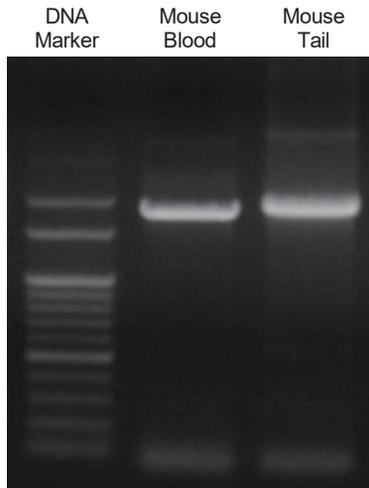
PCR analysis of genomic DNA extracted from 11 plant leaf tissues and seeds of 4 plant species with the P18S primer set.

Related Products

- | | |
|-----------------------------------|------|
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| • YEAtaq DNA Polymerase | 2-1 |
| • RealStart DNA Polymerase Premix | 2-2 |
| • EZtime Real-Time PCR Premix | 2-3 |
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Mouse Tissue

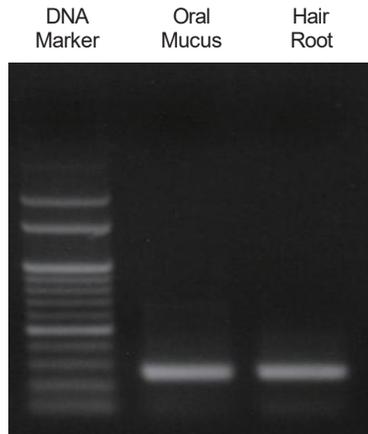
PCR analysis of genomic DNA extracted from ICR mouse blood and tail with a U18S primer set.



* Put 2 μ l of mouse blood and a small piece (~ 1 mm³) of mouse tail tip, respectively, into microcentrifuge tubes then extracted with UniversAll™ buffer.

Human Tissue

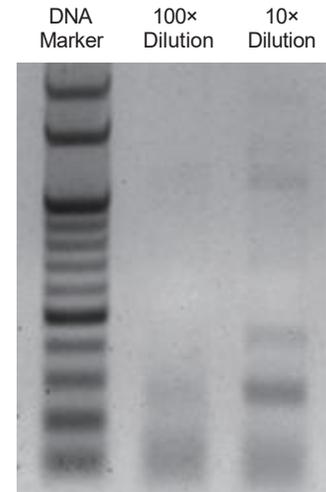
PCR analysis of genomic DNA extracted from human oral mucosal cells and human hair with a β -GBN primer set.



* Placed oral mucosal cells and three hair root ends into microcentrifuge tubes then extracted with UniversAll™ buffer.

Paraffin Sample

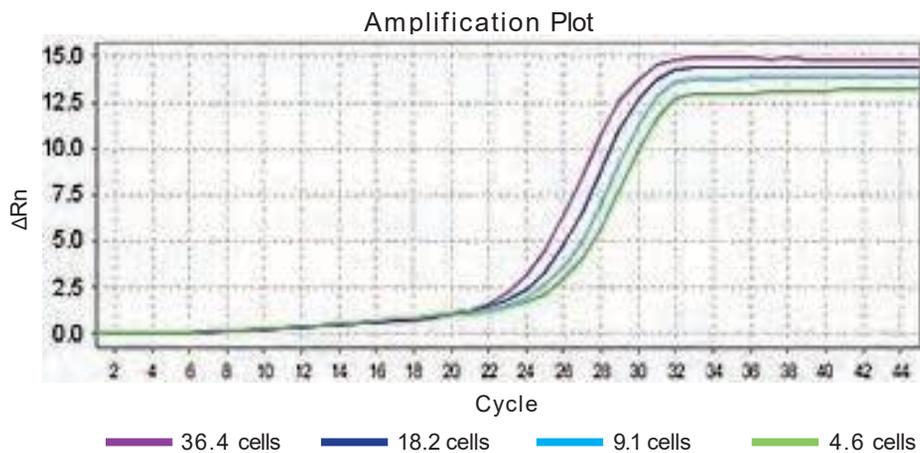
Genomic DNA extracted from paraffin embedded mouse liver with a β -GBN primer set.



* Samples were cut into small pieces, pre-washed with xylene and alcohol then extracted with UniversAll™ buffer.

Real-time Quantitative PCR

Real-time quantitative PCR analysis of *E. coli* plasmid DNA extracted by UniversAll™ extraction buffer.



* As few as 4.6 cells in the sample can be detected easily.

DR Keeper Nucleic Acid Stabilizer



RT

FYU201-100ML

DR Keeper Nucleic Acid Stabilizer Solution

Description

DR Keeper Nucleic Acid Stabilizer is a non-toxic tissue and cell storage solution to stabilize and protect DNA/RNA for later isolation or shipping without refrigeration. It penetrates into tissue pieces or cells rapidly to inactivate RNase and keeps RNA intact. It allows recovery of high quality and quantity of RNA from tissue pieces or cells stored in DR Keeper Nucleic Acid Stabilizer for 1 month at 4°C, 1 week at 25°C or 1 day at 37°C. For long-term storage, tissue pieces or cells can be stored at -20°C. The yellow color of DR Keeper Nucleic Acid Stabilizer indicates the mild acidic condition which protects RNA from nucleophilic attack.

Features

- Preserves RNA at ambient temperatures without degradation
- Penetrates into tissue pieces or cells to stabilize RNA immediately
- Excellent for field sample collection
- Non-toxic and environmental friendly
- Minimizes the usage of liquid N₂
- Eliminates the need to store RNA in freezers or to ship by dry ice
- Compatible with most downstream applications (ex: TRIzol® reagent)

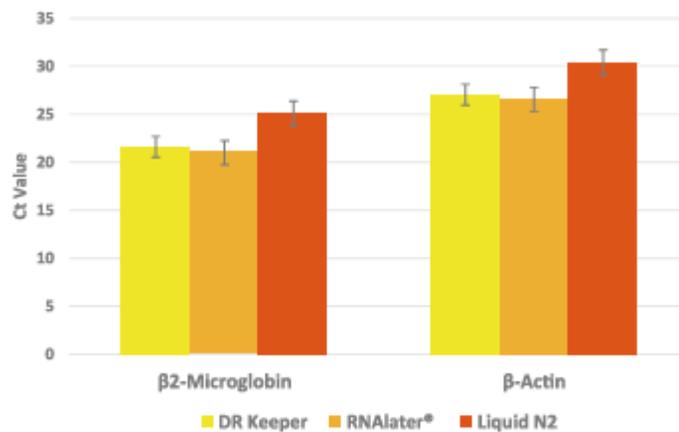
Applications

- DNA preservation of animal tissues, plant tissues, cells
- RNA preservation of animal tissues, plant tissues, cells

Quality Control

DR Keeper Nucleic Acid Stabilizer Solution undergoes quality assurance test to verify that its composition is invariant from lot to lot. RNase digestion assay is performed to ensure its function to protect RNA from degradation.

Results



Q-PCR Result of Mouse Liver's House Keeping Genes stored in different RNA Preservation Reagent for 7 Days in 37°C

Related Products

- | | |
|-----------------------------------|------|
| • YEA Ladder DNA Markers | 1-12 |
| • YEAtaq DNA Polymerase | 2-1 |
| • RealStart DNA Polymerase Premix | 2-2 |
| • EZtime Real-Time PCR Premix | 2-3 |
| • EZtime LAMP Premix (2X) | 2-9 |
| • Deoxynucleotides (dNTPs) | 2-10 |

HiYield Plasmid Mini Kit 3.0

Description

HiYield Plasmid Mini Kit 3.0 is specially designed for rapid isolation of plasmid or cosmid DNA from 1-5 ml of bacterial cultured cells. As high as 40 µg of high quality plasmid DNA can be purified in less than 30 minutes and is ready for restriction digestion, ligation, PCR, and sequencing reaction.

No phenol extraction or alcohol precipitation is required in this protocol. In the process, clear and extra pure cell lysate with minimal genomic DNA and RNA contaminants can be obtained through the modified alkaline lysis method and RNase treatment. In the presence of a chaotropic salt, the plasmid DNA within the lysate will then bind to the glass fiber matrix equipped in the spin column. The contaminants are washed away with an ethanol-containing wash buffer. Finally, the purified plasmid DNA is eluted by a low salt elution buffer or distilled water. Typical yields of high-purity are 20-40 µg for high-copy number plasmids or 3-10 µg for low-copy number plasmids.

Features

- **Sample** : 1-5 ml of bacterial cells
- **Format** : spin column (centrifuge)
- **Yield** : up to 40 µg of plasmid/cosmid DNA
- **Operation time** : up to 30 minutes
- **Elution volume** : 30-50 µl

Applications

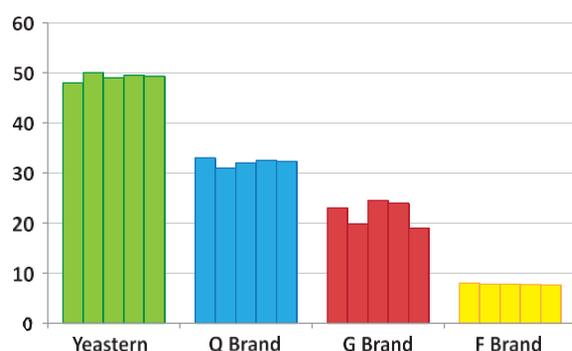
Restriction enzyme digestion, library screening, ligation, PCR, transformation/sequencing reactions

Quality Control

The quality of HiYield Plasmid Mini Kit 3.0 is tested on a lot-to-lot basis. The kit is tested by isolation of plasmid DNA from 5 ml culture of *E. coli* DH5α transformed with the plasmid pGAD424 ($A_{600} > 2$ units/ml). More than 40 µg of plasmid DNA should be obtained. One µg of the purified product is also tested for restriction enzyme digested with *EcoR* I followed by agarose gel analysis.

Results

Performance of Yeastern Biotech's
HiYield Plasmid Mini Kit 3.0



Sample : *E. coli* DH5α

Plasmid : GAD424 (6.6 kb)

Preparation : 16 hours overnight LB culture, 5 ml for each preparation



RT

FYG011-100P (100 preps)

| | |
|--------------------|---------|
| PN1 Buffer (PS1) | 25 mL |
| PN2 Buffer (PS2) | 25 mL |
| PN3 Buffer (PS3) | 35 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| RNase A (50 mg/mL) | 50 µl |
| PN Column (PS) | 100 pcs |
| Collection Tube | 100 pcs |

FYG011-300P (300 preps)

| | |
|--------------------|-----------|
| PN1 Buffer (PS1) | 65 mL |
| PN2 Buffer (PS2) | 65 mL |
| PN3 Buffer (PS3) | 95 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| RNase A (50 mg/mL) | 150 µl |
| PN Column (PS) | 300 pcs |
| Collection Tube | 300 pcs |

Related Products

| | |
|-----------------------------------|------|
| • YEA Ladder DNA Markers | 1-12 |
| • YEAtaq DNA Polymerase | 2-1 |
| • RealStart DNA Polymerase Premix | 2-2 |
| • EZtime Real-Time PCR Premix | 2-3 |
| • Deoxynucleotides (dNTPs) | 2-10 |
| • ECOS™ Competent cells | 3-1 |
| • T&A Cloning Kit | 3-14 |

HiYield Gel/PCR DNA Fragments Extraction Kit 2.0



RT

FYG206-100 (100 preps)

| | |
|-----------------|---------|
| GP Buffer | 60 ml |
| W1 Buffer | 45 ml |
| W2 Buffer | 15 ml |
| Elution Buffer | 10 ml |
| GP Column | 100 pcs |
| Collection Tube | 100 pcs |

FYG206-300 (300 preps)

| | |
|-----------------|----------|
| GP Buffer | 80 ml x2 |
| W1 Buffer | 125 ml |
| W2 Buffer | 25 ml x2 |
| Elution Buffer | 30 ml |
| GP Column | 300 pcs |
| Collection Tube | 300 pcs |

Description

The HiYield Gel/PCR DNA Fragments Extraction Kit 2.0 is designed to recover or concentrate DNA fragments (50 bp-10 kb) from agarose gels in 20 mins, PCR or other enzymatic reactions. The unique dual purpose application and high yield mini columns make this kit valuable. The method uses a chaotropic salt, guanidine thiocyanate to dissolve agarose gel and denature enzymes. DNA fragments in chaotropic salt solution bind to the glass fiber matrix of the spin column. Following washing off contaminants, the purified DNA fragments (Effective Binding Capacity Approx: 20 ug) are eluted by addition of low salt elution buffer or water. Salts, enzymes and unincorporated nucleotides are effectively removed from reaction mixtures without phenol extraction or alcohol precipitation.

Features

- **Recovery** : Up to 95%
- **Operation time** : within 20 minutes

Sample

100 µl PCR Product, 300 mg of Agarose Gel

Applications

Fluorescent/radioactive sequencing, PCR, restriction enzyme digestion, DNA labeling and ligation

Genomic DNA Extraction Mini Kit (Blood and Urine)



RT

FYG109-100 (100 preps)

| | |
|-----------------|---------|
| BU1 Buffer(B1) | 100 ml |
| BU2 Buffer(B2) | 35 ml |
| BU3 Buffer(BC) | 45 ml |
| W1 Buffer | 45 ml |
| W2 Buffer | 15 ml |
| Elution Buffer | 10 ml |
| BU Column(BZ) | 100 pcs |
| Collection Tube | 100 pcs |

Description

The Genomic DNA Extraction Mini Kit (Blood and Urine) is designed for rapid extraction of pure genomic DNA from Bacteria and fungus cell. Efficiently remove cellular debris and inhibitors, this kit using column-type tube in purification process through three simple steps of binding, washing and then elution for the safe and convenient extraction of high-purity genomic DNA. The entire process can be completed in less than 1 hour without phenol/chloroform, and the final product can be used in PCR or other downstream experiments.

Features

- **Format** : spin column (centrifuge)
- **Yield** : up to 50 µg

Sample

310⁹ Bacteria, 5 x 10⁷ Fungus cells

Applications

PCR, AFLP/PADP, RFLP, Southern blot, real-time PCR

Genomic DNA Extraction Kit (Tissue) 2.0

Description

The Genomic DNA Extraction Kit (Tissue) 2.0 is designed for rapid extraction of pure genomic DNA from animal tissue or paraffin-embedded tissue. Efficiently remove cellular debris and inhibitors, this kit using column-type tube in purification process through three simple steps of binding, washing and then elution for the safe and convenient extraction of high-purity genomic DNA. The entire process can be completed in less than 1 hour without phenol/chloroform, and the final product can be used in PCR or other downstream experiments

Features

- **Format** : spin column (centrifuge)
- **Yield** : up to 50 µg

Sample

330 mg of fresh animal tissue, 25 mg of paraffin-embedded tissue

Applications

PCR, AFLP/PADP, RFLP, Southern blot, real-time PCR



RT

FYG111-100 (100 preps)

| | |
|-----------------|---------|
| T1 Buffer (T1) | 35 mL |
| T12 Buffer (T2) | 12 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| TI Column (GZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG111-300 (300 preps)

| | |
|-----------------|-----------|
| T1 Buffer (T1) | 95 mL |
| T12 Buffer (T2) | 35 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| TI Column (GZ) | 300 pcs |
| Collection Tube | 300 pcs |

Genomic DNA Extraction Kit (Plant) 2.0

Description

The Genomic DNA Extraction Kit (Plant) 2.0 is designed for rapid extraction of pure genomic DNA from plant tissue or dry plant tissue. Efficiently remove cellular debris and inhibitors, this kit using column-type tube in purification process through three simple steps of binding, washing and then elution for the safe and convenient extraction of high-purity genomic DNA. The entire process can be completed in less than 1 hour without phenol/chloroform, and the final product can be used in PCR or other downstream experiments

Features

- **Format** : spin column (centrifuge)
- **Yield** : up to 50 µg

Sample

100 mg of fresh plant tissue, 50 mg of dry plant tissue

Applications

PCR, AFLP/PADP, RFLP, Southern blot, real-time PCR



RT

FYG112-100 (100 preps)

| | |
|-----------------|---------|
| PL Buffer (PZ) | 55 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| PL Column (GZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG112-300 (300 preps)

| | |
|-----------------|-------------|
| PL Buffer (PZ) | 125 + 30 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| PL Column (GZ) | 300 pcs |
| Collection Tube | 300 pcs |

Genomic DNA Extraction Kit (Bacteria/Fungi) 2.0



RT

FYG115-100 (100 preps)

| | |
|-----------------|---------|
| BF1 Buffer (N1) | 20 mL |
| BF2 Buffer (N2) | 35 mL |
| BF3 Buffer (N3) | 45 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| BF Column (GZ) | 100 pcs |
| Collection Tube | 100 pcs |

Description

The Genomic DNA Extraction Kit (Bacteria/Fungi) 2.0 is designed for rapid extraction of pure genomic DNA from Bacteria and fungus cell. Efficiently remove cellular debris and inhibitors, this kit using column-type tube in purification process through three simple steps of binding, washing and then elution for the safe and convenient extraction of high-purity genomic DNA. The entire process can be completed in less than 1 hour without phenol/chloroform, and the final product can be used in PCR or other downstream experiments.

Features

- **Format** : spin column (centrifuge)
- **Yield** : up to 50 µg

Sample

10⁹ Bacteria, 5 x 10⁷ Fungus cells

Applications

PCR, AFLP/PADP, RFLP, Southern blot, real-time PCR

Genomic DNA Extraction kit (Cultured Cell) 2.0



RT

FYG117-100 (100 preps)

| | |
|-----------------|---------|
| CC1 Buffer (N1) | 20 mL |
| CC2 Buffer (N2) | 35 mL |
| CC3 Buffer (N3) | 45 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| CC Column (GZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG117-300 (300 preps)

| | |
|-----------------|-----------|
| CC1 Buffer (N1) | 20 mL |
| CC2 Buffer (N2) | 95 mL |
| CC3 Buffer (N3) | 125 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| CC Column (GZ) | 300 pcs |
| Collection Tube | 300 pcs |

Description

The Genomic DNA Extraction kit (Cultured Cell) 2.0 is designed for rapid extraction of pure genomic DNA from Cultured Cell. Efficiently remove cellular debris and inhibitors, this kit using column-type tube in purification process through three simple steps of binding, washing and then elution for the safe and convenient extraction of high-purity genomic DNA. The entire process can be completed in less than 1 hour without phenol/chloroform, and the final product can be used in PCR or other downstream experiments.

Features

- **Format** : spin column (centrifuge)
- **Yield** : up to 50 µg

Sample

10⁷ Cultured Cell

Applications

PCR, AFLP/PADP, RFLP, Southern blot, real-time PCR

Total RNA kit (Cell/Blood)

Description

The Total RNA kit (Cell/Blood) is designed specifically for purifying total RNA from fresh whole human blood and cultured cells. Detergents and a chaotropic salt are used to lyse cells and inactivate RNase. RNA in the chaotropic salt is bound to the glass fiber matrix of the spin column and once any contaminants have been removed using the Wash Buffer, the purified total RNA is eluted by the Elution Buffer.

Features

- **Format** : spin column (centrifuge)
- **Yield** : Up to 30 µg

Sample

10^7 x Culture Cells, 300 µl of blood

Applications

RT-PCR, Northern blotting, primer extension, mRNA selection cDNA synthesis



RT

FYG310-100 (100 preps)

| | |
|-------------------|---------|
| CBR1 Buffer (CR1) | 110 mL |
| CBR2 Buffer (CR2) | 45 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| CBR Column (RZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG310-300 (300 preps)

| | |
|-------------------|------------|
| CBR1 Buffer (CR1) | 110 * 3 mL |
| CBR2 Buffer (CR2) | 125 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| CBR Column (RZ) | 300 pcs |
| Collection Tube | 300 pcs |

Total RNA kit (Blood/ Bacteria)

Description

The Total RNA kit (Blood/ Bacteria) is designed specifically for purifying total RNA from fresh whole human blood and Bacteria. Detergents and a chaotropic salt are used to lyse cells and inactivate RNase. RNA in the chaotropic salt is bound to the glass fiber matrix of the spin column and once any contaminants have been removed using the Wash Buffer, the purified total RNA is eluted by the Elution Buffer.

Features

- **Format** : spin column (centrifuge)
- **Yield** : Up to 30 µg

Sample

10^9 x Bacteria Cells, 300 µl of blood

Applications

RT-PCR, Northern blotting, primer extension, mRNA selection cDNA synthesis



RT

FYG306-100 (100 preps)

| | |
|-------------------|---------|
| BFR1 Buffer (BRO) | 20 mL |
| BFR2 Buffer (BRA) | 45 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| BFR Column (RZ) | 100 pcs |
| Collection Tubes | 100 pcs |

FYG306-300 (300 preps)

| | |
|-------------------|-----------|
| BFR1 Buffer (BRO) | 60 mL |
| BFR2 Buffer (BRA) | 65 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| BFR Column (RZ) | 300 pcs |
| Collection Tubes | 300 pcs |

Total RNA Kit (Tissue) 2.0



FYG307-100 (100 preps)

| | |
|-----------------|---------|
| TIR Buffer (TR) | 45 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| TIR Column (RZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG307-300 (300 preps)

| | |
|-----------------|-----------|
| TIR Buffer (TR) | 125 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| TIR Column (RZ) | 300 pcs |
| Collection Tube | 300 pcs |

Description

The Total RNA Kit (Tissue) 2.0 is designed specifically for purifying total RNA from fresh animal tissue or paraffin-embedded tissue. Detergents and a chaotropic salt are used to lyse cells and inactivate RNase. RNA in the chaotropic salt is bound to the glass fiber matrix of the spin column and once any contaminants have been removed using the Wash Buffer, the purified total RNA is eluted by the Elution Buffer.

Features

- **Format** : spin column (centrifuge)
- **Yield** : Up to 30 µg

Sample

30 mg of fresh animal tissue, 25 mg of paraffin-embedded tissue

Applications

RT-PCR, Northern blotting, primer extension, mRNA selection cDNA synthesis

Total RNA Kit (Plant) 2.0



FYG308-100 (100 preps)

| | |
|-----------------|---------|
| PLR Buffer (PR) | 110 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| PLR Column (RZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG308-300 (300 preps)

| | |
|-----------------|------------|
| PLR Buffer (PR) | 105 * 3 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| PLR Column (RZ) | 300 pcs |
| Collection Tube | 300 pcs |

Description

The Total RNA Kit (Plant) 2.0 is designed specifically for purifying total RNA from plant tissue or dry plant tissue. Detergents and a chaotropic salt are used to lyse cells and inactivate RNase. RNA in the chaotropic salt is bound to the glass fiber matrix of the spin column and once any contaminants have been removed using the Wash Buffer, the purified total RNA is eluted by the Elution Buffer.

Features

- **Format** : spin column (centrifuge)
- **Yield** : Up to 30 µg

Sample

100 mg of plant Tissue, 25 mg of dry plant Tissue

Applications

RT-PCR, Northern blotting, primer extension, mRNA selection cDNA synthesis

miRNA Isolation Kit 2.0

Description

The miRNA Isolation Kit 2.0 is designed for purifying micro RNAs (miRNAs) and other small cellular RNAs from whole blood, cultured cells, bacteria (Gram +/-), fungus cells and tissue. Standard protocols for isolating total RNA and mRNA are not optimized for isolation of small RNA molecules and result in the loss of substantial amounts of miRNAs and other small RNA. In addition, removal of the predominantly larger RNAs is required for accurate analysis of miRNA expression by qPCR or microarray analysis. This kit ensures purification of small RNA with minimal contamination from large RNA molecules or genomic DNA. In the presence of a chaotropic salt, and various ethanol concentrations in the solvent, RNA molecules (of various sizes) are selectively bound to the glass fiber matrix.

Features

- **Format** : spin column (centrifuge)
- **Yield** : Up to 20 µg

Sample

500 µl Whole Blood, 10⁶ Cultured Cells, 10⁸ Bacteria (Gram +/-), 5x10⁶ fungus cells, 10 mg Tissue

Applications

Northern blot analysis, quantitative, real-time RT-PCR, microarray analysis



RT

FYG309-050 (50 preps)

| | |
|--------------------|----------------|
| PD1 Buffer (PM1) | 85 mL |
| PD2 Buffer (PM2) | 85 mL |
| PD3 Buffer (PM3) | 125 mL |
| W1 Buffer | 125 mL + 40 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 50 mL |
| RNase A (50 mg/mL) | 200 µl |
| 3M Sodium Acetate | 4 mL * 2 |
| PD Column (PM) | 20 pcs |

Viral Nucleic Acid Extraction Kit 2.0

Description

Viral Nucleic Acid Extraction Kit 2.0 is specially designed for purification of viral RNA or DNA from cell-free samples. With the extraction method included, DNA/RNA viruses are lysed quickly and efficiently by the lysis buffer which is a highly concentrated solution of a chaotropic salt. The lysis buffer and ethanol create appropriate conditions for the binding of nucleic acids to the glass fiber matrix of the blood viral DNA/RNA binding column. Contaminations like salts, metabolites and soluble macromolecular cellular components are removed in the wash steps. The nucleic acids can be eluted in low salt buffer or water and are ready-to-use in subsequent reactions.

Features

- **Format** : spin column (centrifuge)
- **Yield** : Up to 20 µg

Sample

serum, plasma, cell-culture supernatants, other cell-free body fluids

Applications

RT-PCR, PCR, Real-time PCR, Automated fluorescent DNA sequencing, Enzymatic reactions



RT

FYG404-050 (50 preps)

| | |
|------------------|---------|
| VN1 Buffer (VN1) | 45 mL |
| VN2 Buffer (VN2) | 6 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| VN Column (VZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG404-300 (300 preps)

| | |
|------------------|-----------|
| VN1 Buffer (VN1) | 125 mL |
| VN2 Buffer (VN2) | 16 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| VN Column (VZ) | 300 pcs |
| Collection Tube | 300 pcs |

Genomic DNA Extraction Kit (Stool) 2.0



RT

FYG113-100 (100 preps)

| | |
|------------------|---------|
| SS1 Buffer (ST1) | 35 mL |
| SS2 Buffer (ST2) | 12 mL |
| W1 Buffer | 45 mL |
| W2 Buffer | 15 mL |
| Elution Buffer | 10 mL |
| Proteinase K | 40 mg |
| Bead Tube | 100 pcs |
| SS Column (GZ) | 100 pcs |
| Collection Tube | 100 pcs |

FYG113-300 (300 preps)

| | |
|------------------|-----------|
| SS1 Buffer (ST1) | 95 mL |
| SS2 Buffer (ST2) | 35 mL |
| W1 Buffer | 125 mL |
| W2 Buffer | 25 * 2 mL |
| Elution Buffer | 30 mL |
| Proteinase K | 40 * 3 mg |
| Bead Tube | 300 pcs |
| SS Column (GZ) | 300 pcs |
| Collection Tube | 300 pcs |

Description

The Genomic DNA Extraction Kit (Stool) 2.0 is designed for rapid extraction of pure genomic DNA from stool. Efficiently remove cellular debris and inhibitors, this kit using column-type tube in purification process through three simple steps of binding, washing and then elution for the safe and convenient extraction of high-purity genomic DNA. The entire process can be completed in less than 1 hour without phenol/ chloroform, and the final product can be used in PCR or other downstream experiments

Features

- **Format** : spin column (centrifuge)
- **Yield** : up to 50 µg

Sample

30 mg of stool sample

Applications

PCR, AFLP/PADP, RFLP, Southern blot, real-time PCR

YEA Ladder DNA Markers

Description

The Yeastern Biotech's 1 kb YEA Ladder DNA Marker IV (FYD011-001) contains 13 discrete fragments ranging from 250 bp to 12 kb. The 100 bp YEA Ladder DNA Marker IV (FYD012-001) contains 15 discrete fragments ranging from 100 bp to 5000 bp. It is supplied in a ready-to-use format containing blue tracking dyes and mixed at varied concentrations. It allows sizing and concentration estimation of DNA fragments on agarose gels generated by PCR or restriction digestion.

Features

- Sharp and clear banding patterns
- Mass marker bands for easy DNA quantification
- Accurate DNA migration on agarose gel

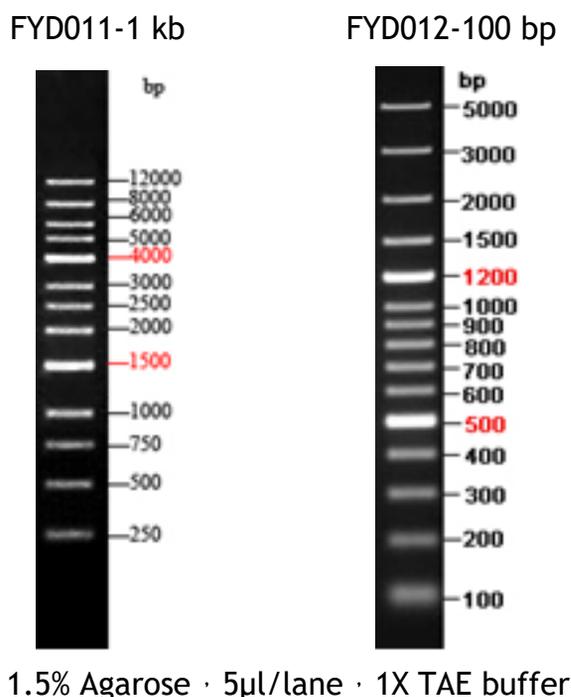
Applications

- Locating DNA of interest
- DNA quantification
- DNA tracking

Quality Control

5 μ l of YEA Ladder DNA Markers provide expected numbers of bands with accurate molecular weight on agarose gel after running electrophoresis.

Results



FYD011-1ML

1 kb YEA Ladder DNA Marker IV
1ml

FYD012-1ML

100 bp YEA Ladder DNA Marker IV
1ml

Related Products

- | | |
|---------------------------------------|------|
| • YEAtaq DNA Polymerase | 2-1 |
| • RealStart DNA Polymerase Premix | 2-2 |
| • EZtime Real-Time PCR Premix | 2-3 |
| • Deoxy+ HiSpec Reverse Transcriptase | 2-5 |
| • Deoxy+ OneStep RT-PCR Kit | 2-6 |
| • Deoxynucleotides (dNTPs) | 2-10 |
| • T&A Cloning Kit | 3-14 |

EtB“Out” Nucleic Acid Staining Solution



RT

FYD010-200P (200 preps)

EtB“Out” Nucleic Acid Staining Solution (2.0)
1ml

Description

EtB“Out” Nucleic Acid Staining Solution aims to replace traditional EtBr (ethidium bromide) in performing nucleic acid detection in agarose gels. EtBr has long been known as a strong mutagen, however EtB“Out” causes only neglectable mutations in the Ames test.

The sensitivity of EtB“Out” is identical to that of EtBr. Under UV light, EtB“Out” emits green fluorescence when bound to DNA or RNA. EtB“Out” can be excited at 290 nm and 490 nm. The fluorescence emission peak of EtB“Out” when bound to DNA is at 537 nm.

Features

- **Economic** : use only 5 µl in 100 ml of agarose gel
- **Sensitive** : sensitivity is comparable and even better than EtBr, DNA concentration as low as 5 ng can be detected
- **Safe** : non-mutagenic, non-toxic, non carcinogenic
- **Green** : no hazardous waste

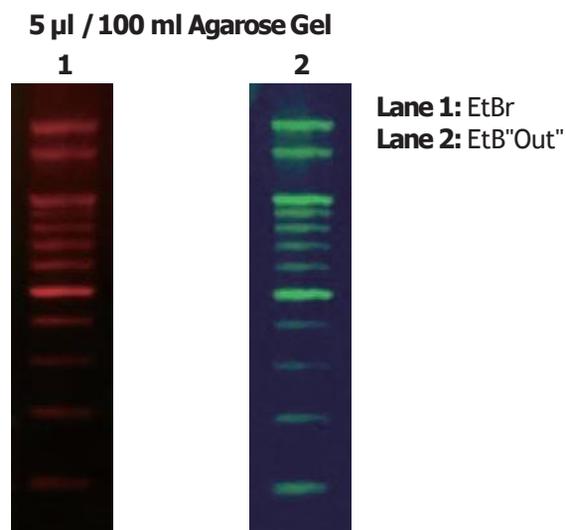
Applications

Nucleic acids detection (dsDNA and ssRNA) within agarose gel after electrophoresis under UV illumination

Quality Assurance

The quality of the EtB“Out” Nucleic Acid Staining Solution is tested on a lot-to-lot basis. Nucleic acids is extracted from tissues and serial diluted. Electrophoresis is performed and the agarose gel is stained with EtB“Out” Nucleic Acid Staining Solution to ensure the performance.

Results



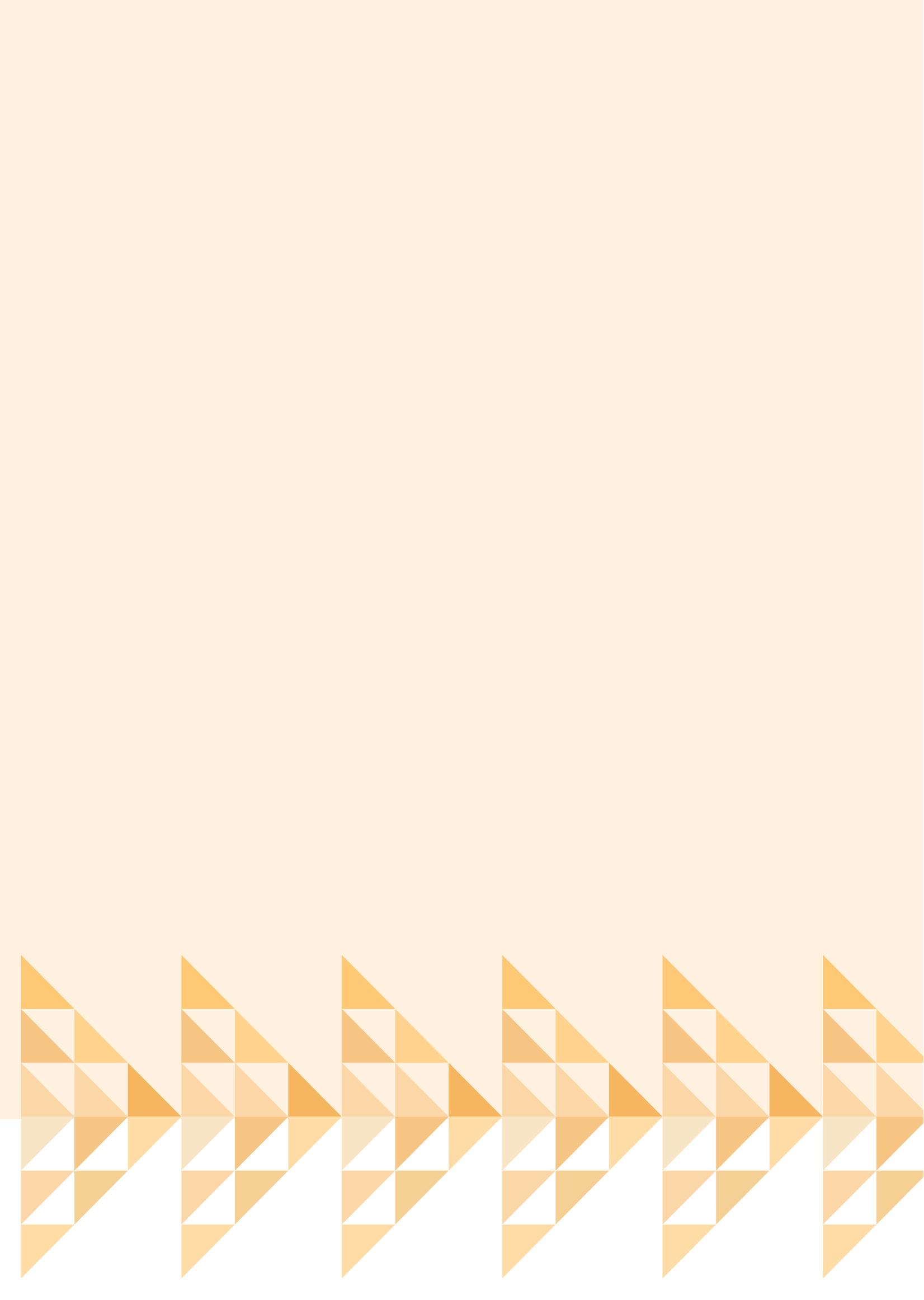
Nucleic staining using EtB“Out” Nucleic Acid Staining Solution.

Two individual agarose gels were prepared with 5 µl of EtBr and EtB“Out” respectively. Staining results were examined under UV illumination. Identical results were observed and EtB“Out” even showed a better staining result than using EtBr.

Related Products

- | | |
|---------------------------------------|------|
| • YEAtaq DNA Polymerase | 2-1 |
| • RealStart DNA Polymerase Premix | 2-2 |
| • EZtime Real-Time PCR Premix | 2-3 |
| • Deoxy+ HiSpec Reverse Transcriptase | 2-5 |
| • Deoxy+ OneStep RT-PCR Kit | 2-6 |
| • Deoxynucleotides (dNTPs) | 2-10 |
| • T&A Cloning Kit | 3-14 |

Note





PCR, RT-PCR & dNTPs

YEAtaq DNA Polymerase



FYT601-500U (500 units)
 YEAtaq DNA Polymerase (5 U/μl) 100 μl
 10× Reaction Buffer 2 ml
 dNTPs Mix (10 mM) 200 μl

FYT611-500U (500 units)
 YEAtaq DNA Polymerase (5 U/μl) 100 μl
 10× Reaction Buffer 2 ml

* The reaction buffer is supplied as a 10× concentrate and should be diluted for use.

Description

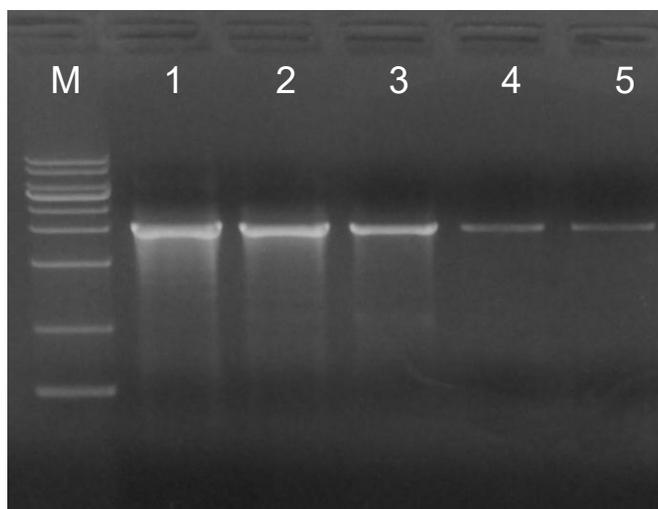
YEAtaq II DNA Polymerase is a thermostable enzyme derived from *Thermus aquaticus* YT-1 strain. The enzyme is in a recombinant form expressed in *E. coli*. It is able to withstand repeated heating to 95°C without significant loss of activity. It possesses both 5'-3' polymerase and exonuclease activity, and has no detectable 3'-5' exonuclease activity. It has a 3' adenylation activity. Thus, the PCR products can be used directly in TA-cloning procedures.

Yeastern Biotech offers YEAtaq DNA Polymerase in two different packages: one with dNTPs mix and the other without.

Applications

- Cloning
- Screening
- Primer extension
- Terminal dA tailing
- Routine PCR amplification of DNA fragments up to 3 Kb
- DNA labeling
- DNA sequencing

Results



Template concentration:
 Lane M : 1kb DNA Marker
 Lane 1 : 1 ng/μl
 Lane 2 : 0.1 ng/μl
 Lane 3 : 0.01 ng/μl
 Lane 4 : 0.001 ng/μl
 Lane 5 : 0.0005 ng/μl

Related Products

- UniversAll™ Tissue Extraction/PCR Kits 1-1
- YEA Ladder DNA Markers 1-12
- Deoxynucleotides (dNTPs) 2-10
- T&A Cloning Kit 3-14

Patterns/Disclaimer

Some applications in which this product can be used may be covered by patents issued and applicable in certain countries. Because purchase of this product does not include a license to perform any patented application, users of this product may be required to obtain a patent license depending upon the particular application and country in which the product is used.

RealStart DNA Polymerase Premix

Description

RealStart DNA Polymerase premix is an ultra-sensitive and convenient PCR premix product. It contains 2× concentrated solution of HotStart DNA polymerase, dNTPs, optimized buffers, and loading dye (optional) needed for PCR. The only step it takes to perform PCR with RealStart DNA Polymerase premix is to add DNA template and primers into the reaction mix. Since special HotStart DNA polymerase in the premix is activated after heating, it greatly reduces non-specific amplification when working with the premix at room temperature.

Features

- **Simple & time-saving** : Just add templates of interest and primers into the RealStart DNA Polymerase Premix.
- **Less Contamination** : Reduce non-specific amplification caused by mispriming events that occur during setup and initial temperature increase.
- **High Sensitivity** : tested in amplification of a single gene copy.
- **Convenient** : An excellent tool when working with high quantities of samples.
- **Sample Size** : work excellent for short DNA templates (size shorter than 600 bp).

Applications

- High throughput hot-start PCR.
- RT-PCR.
- Highly specific amplification of complex genomic and cDNA templates.
- Amplification of low copy DNA targets.
- Generation of PCR products for TA cloning.

Unit Definition

One unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTPs into acid-insoluble material in 30 minutes at 72 °C.

Quality Control

- Nuclease activity is not detected after incubation of 1 µg λ /Hind III DNA with 5 units of RealStart DNA polymerase in 50 µl reaction buffer for 18 hours at 37 °C.
- The absence of endo-, exodeoxyribonucleases and ribonucleases is confirmed by appropriate tests. Functional test is performed by PCR.



-20°C

FYT101-100P (100 preps)

| | |
|-----------------------------|----------|
| 2× RealStart DNA Polymerase | 1.25 ml |
| Premix (w/o loading dye) | 0.1 U/µl |

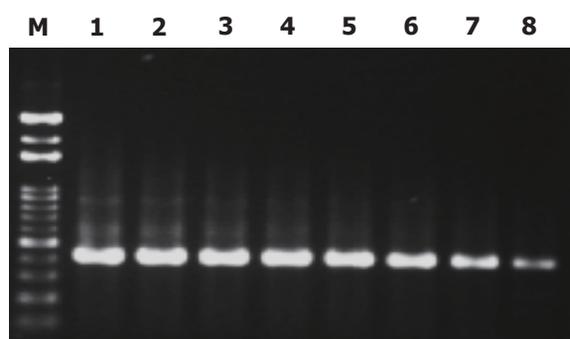
Premix contains:

- Hotstart Taq DNA polymerase
- dNTPs mix (including dATP, dCTP, dGTP, dTTP)
- 7.5 mM MgCl₂

Related Products

- | | |
|--|------|
| • UniversAll™ Tissue Extraction/PCR Kits | 1-1 |
| • YEA Ladder DNA Markers | 1-12 |
| • T&A Cloning Kit | 3-14 |

Results



High sensitivity for low copy genes!

Template: pUC18
1 mg pUC18 DNA = 3.4 x 10¹¹ molecules

| | |
|--|--|
| Lane M: 1 kb DNA Marker (#FYD001-500UL) | |
| Lane 1: 3 x 10 ⁷ molecules | Lane 2: 3 x 10 ⁶ molecules |
| Lane 3: 3 x 10 ⁵ molecules | Lane 4: 3 x 10 ⁴ molecules |
| Lane 5: 3 x 10 ³ molecules | Lane 6: 3 x 10 ² molecules |
| Lane 7: 3 x 10 ¹ molecules | Lane 8: 3 molecules |

EZtime Real-Time PCR Premix(SYBR Green)



FYT104-100P (100 preps)

EZtime Fast Real-Time PCR Premix (2×, SYBR Green I, ROX) 1 ml

FYT104-400P (400 preps)

EZtime Fast Real-Time PCR Premix (2×, SYBR Green I, ROX) 1 ml x 4

Premix contains:

- Hotstart Taq DNA polymerase
- SYBR Green real-time PCR Buffer
- dNTP mix including dATP, dCTP, dGTP, dTTP
- 5 mM MgCl₂

Description

The EZtime Real-Time PCR Premix (**# FYT104-100P**) is a ready-to-use, 2× concentrated PCR premix, including Hotstart Taq, SYBR Green I, optimized reaction buffer and nucleotides (**# FYT104-100P with ROX**) for running quantitative real-time PCR, including qPCR and 2-step qRT-PCR, in the SYBR Green I detection format. SYBR Green Premix provides high sensitivity, wide dynamic range and reproducibility for quantification.

Features

- **Specificity** : Hotstart Taq and the optimized buffer eliminates non-specific amplification and formation of primer dimers
- **Sensitivity** : detects low copy number targets
- **Wide linear range** : accurate quantification across 9 orders of magnitude
- **Reproducibility and convenience** : ready-to-use 2× master mix minimizes pipetting error and reduces set-up time

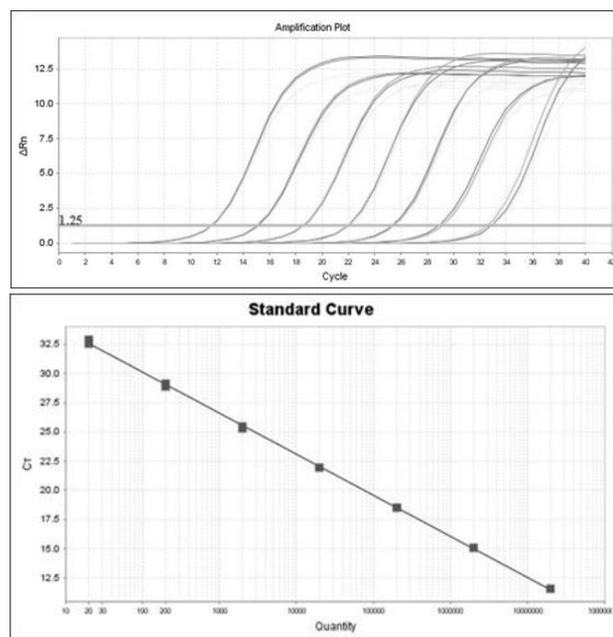
Applications

- Quantitative real-time PCR
- Quantitative 2-step RT-PCR
- Quick and accurate detection and quantification of target gene through real-time PCR

Quality Control

- Error<0.01, 2.1>Efficiency>1.90, ΔCP<1
- The absence of endo-, exodeoxyribonucleases and ribonucleases is confirmed by appropriate quality tests. Functionally tested in qPCR for specificity, sensitivity and reproducibility using serial dilutions of control genomic DNA template.

Results



Primers were designed to amplify a specific region of YB T&A™ Vector that has been serially diluted to different concentrations.

Related Products

- UniversAll™ Tissue Extraction/PCR Kits 1-1
- YEA Ladder DNA Markers 1-12

EZtime Real-Time PCR Premix(TaqMan)

Description

EZtime Fast Real-Time PCR Premix for TaqMan® Probe (**#FYT110-100P/400P**) is a ready-to-use, 2× concentrated premix reagent, containing all components except primers, probe, and template. It is formulated with a novel hot-start Taq DNA polymerase, which is capable of catalyzing DNA amplification in a fast PCR mode. This special blend greatly shortens the running time of real-time quantitative PCR by around 1 hour when compared to traditional qPCR. In addition, it precisely meets current researchers' needs for performing gene detection (qPCR) and quantification of gene expression (2-step qRT-PCR) in a high speed and/or high-throughput manner in addition to those basic requirements of high sensitivity, wide dynamic range, and good reproducibility.

Features

- **Fast** : shortens the running time of real-time quantitative PCR by around 1 hour
- **Specificity** : Hotstart Taq and the optimized buffer eliminates non-specific amplification and formation of primer dimers
- **Sensitivity** : detects low copy number targets
- **Wide linear range** : accurate quantification across 9 orders of magnitude
- **Reproducibility and convenience** : ready-to-use 2× master mix minimizes pipetting error and reduces set-up time

Applications

- Quantitative real-time PCR
- Quantitative 2-step RT-PCR
- Quick and accurate detection and quantification of target gene through real-time PCR

Quality Control

- Error<0.01, 2.1>Efficiency>1.90, ΔCP<1
- The absence of endo-, exodeoxyribonucleases and ribonucleases is confirmed by appropriate quality tests. Functionally tested in qPCR for specificity, sensitivity and reproducibility using serial dilutions of control genomic DNA template.



-20°C

FYT110-100P (100 preps)

EZtime Fast Real-Time PCR Premix (2×, TaqMan, ROX) 1 ml

FYT110-400P (400 preps)

EZtime Fast Real-Time PCR Premix (2×, TaqMan, ROX) 1 ml x 4

Premix contains:

- Hotstart Taq DNA polymerase
- TaqMan real-time PCR Buffer
- dNTP mix including dATP, dCTP, dGTP, dTTP
- 5 mM MgCl₂

Related Products

- UniversAll™ Tissue Extraction/PCR Kits 1-1
- YEA Ladder DNA Markers 1-12

Deoxy+ HiSpec Reverse Transcriptase



-20°C

FYT501-100R (100 rxns)

| | |
|---|------------|
| Deoxy+ HiSpec Reverse Transcriptase (20000 U) | 50 µl x 2 |
| 2× Deoxy+ RT Premix* | 0.5 ml x 2 |

(* 2× Deoxy+ RT Premix contains RT buffer, 0.1 M DTT and 10 mM dNTPs)

Description

Deoxy+ HiSpec Reverse Transcriptase (RT) is genetically engineered by introducing of point mutations to MMLV RT that increase half-life, reduce RNase activity and increase thermal stability. Those designed mutations lead to increased specificity of Deoxy+ HiSpec RT and the highest cDNA yield of all RTs. It is ideal for RT-PCR of a specific gene or generating cDNA from total or poly (A)+ RNA samples. It synthesizes a complementary DNA strand from total RNA, mRNA, or an RNA:DNA hybrid.

Features

- Half life of 100 minutes at 50°C for the highest cDNA yields
- Reduced RNase H activity for more full-length cDNA
- Full activity at 50°C for increased specificity with GSP
- Ability to increase RT units without inhibiting subsequent PCR

Applications

- Synthesis of first-strand cDNA
- Array labeling
- cDNA libraries
- RT-PCR, primer extension, and 3' and 5' RACE

Unit Definition

One unit incorporates 1 nmole of dTTP into acid precipitable material in 10 minutes at 37°C using poly(A)-oligo(dT) as template primer.

Quality Control

This product has passed the following quality control assays: SDS-polyacrylamide gel analysis for purity; functional absence of endodeoxyribonuclease, 3' and 5' exodeoxyribonuclease, and ribonuclease activities; yield and length of cDNA product.

Related Products

| | |
|--|------|
| • UniversAll™ Tissue Extraction/PCR Kits | 1-1 |
| • YEA Ladder DNA Markers | 1-12 |
| • T&A Cloning Kit | 3-14 |

Deoxy+ OneStep RT-PCR Kit

Description

The Deoxy+ OneStep RT-PCR Kit is a ready-to-use master mix, which eliminates the need for optimization of reaction and cycling conditions for one-step RT-PCR. The reaction can be prepared by simply adding template RNA and primers to the master mix. The use of Yeastern's Hotstart DNA polymerase and Deoxy+ HiSpec RT enables reliable real-time RT-PCR quantification on any real-time PCR machines. Since it is a one-tube reaction, the procedure makes high-throughput analysis possible.

After reverse transcription, reactions are heated to 95°C for 10 minutes to inactivate the reverse transcriptase and simultaneously activate HotStart Taq DNA polymerase. This hot start to the PCR eliminates any nonspecific amplification products such as primer-dimers and reduces background smear, ensuring highly sensitive and reproducible RT-PCR.

Features

- Fast and easy one-tube setup
- One-step RT-PCR of any RNA template without optimization
- Unique enzyme mix for high specificity and sensitivity
- Optimized reverse-transcription and amplification buffer

Applications

- Onestep RT-PCR
- Gene-expression analysis

Quality Control

The performance of Deoxy+ OneStep RT-PCR Kit is tested in an RT reaction using human embryonic kidney cell lysate with primer d(T)20. The sensitivity of the kit is verified by the detection of B2M transcript after 40 cycles. The length of cDNA achieved is verified by detection of a 248 bp by DNA agarose gel and DNA sequencing.



-20°C

FYT503-50P (50 preps)

| | |
|------------------------------------|----------|
| 2× Deoxy+ OneStep RT PCR Premix | 0.625 ml |
| Sterilized ddH ₂ O | 1 ml |

FYT503-100P (100 preps)

| | |
|------------------------------------|--------------|
| 2× Deoxy+ OneStep RT PCR Premix | 0.625 ml x 2 |
| Sterilized ddH ₂ O | 1 ml x 2 |

Related Products

| | |
|--|------|
| • UniversAll™ Tissue Extraction/PCR Kits | 1-1 |
| • YEA Ladder DNA Markers | 1-12 |
| • T&A Cloning Kit | 3-14 |

Deoxy+ Real-time SYBR Green RT-PCR Kit



FYT504-50P (50 preps)

2× Deoxy+ Real-time SYBR Green RT PCR Premix, ROX 0.625 ml

Sterilized ddH₂O 1 ml

FYT504-100P (100 preps)

2× Deoxy+ Real-time SYBR Green RT PCR Premix, ROX 0.625 ml x 2

Sterilized ddH₂O 1 ml x 2

Description

Deoxy+ Real-time SYBR Green RT-PCR system provides users with a rapid and simple way to quantify the expression of gene of interest based on Real-time PCR system containing SYBR Green. Yeastern's Deoxy+ HiSpec RT, Hotstart DNA polymerase and all the components for Real-time SYBR Green RT-PCR are skillfully mixed within a single tube. Unique buffer system allows highly specific quantification by preventing the formation of nonspecific products and primer-dimers.

Features

- Ready-to-use master mix to allow faster setup, maximize throughput and reduce the contamination risk of real-time RT-PCR analysis
- Detection of even low copy numbers
- Excellent sensitivity and easy optimization with SYBR Green

Applications

- Validation of siRNA-mediated gene knockdown
- Detection of gene regulation

Quality Control

The performance of Deoxy+ Real-time SYBR Green RT-PCR Kit is tested in an RT reaction using human embryonic kidney cell lysate with primer d(T)₂₀. The sensitivity of the kit is verified by the detection of B2M transcript after 40 cycles. The length of cDNA achieved is verified by detection of a 248 bp by DNA agarose gel and DNA sequencing.

Related Products

- UniversAll™ Tissue Extraction/PCR Kits 1-1
- T&A Cloning Kit 3-14

Deoxy+ Real-time TaqMan RT-PCR Kit

Description

TaqMan detection system uses a designed fluorogenic probe to detect target PCR products during real-time PCR cycle. Fluorescent is detected when the quencher of probe is removed during the PCR extension cycle. Deoxy+ Real-time TaqMan RT-PCR Kit complete reverse transcription and TaqMan Real-time PCR in a single tube. Yeastern's Deoxy+ HiSpec RT, Hotstart DNA polymerase and all the components necessary for performing TaqMan Real-time PCR from RNA template are specially prepared to ensure high specificity and high sensitivity.

Features

- Ready-to-use master mix to allow faster setup, maximize throughput and reduce the contamination risk of real-time RT-PCR analysis
- Detection of even low copy numbers
- Excellent specificity and reproducibility with better signal to noise ratio
- Higher PCR efficiency

Applications

- Validation of siRNA-mediated gene knockdown
- Detection of multiple gene expressions

Quality Control

The performance of Deoxy+ Real-time TaqMan RT-PCR kit is tested in an RT reaction using human embryonic kidney cell lysate with primer d(T)₂₀. The sensitivity of the kit is verified by the detection of B2M transcript after 40 cycles. The length of cDNA achieved is verified by detection of a 248 bp by DNA agarose gel and DNA sequencing.



-20°C

FYT505-50P (50 preps)

Deoxy+ Real-time Taqman RT PCR Premix, ROX 0.625 ml

Sterilized ddH₂O 1 ml

FYT505-100P (100 preps)

Deoxy+ Real-time Taqman RT PCR Premix, ROX 0.625 ml x 2

Sterilized ddH₂O 1 ml x 2

Related Products

- UniversAll™ Tissue Extraction/PCR Kits 1-1
- T&A Cloning Kit 3-14

EZtime LAMP Premix (2X)



FYT701-50R (50 rxns)

| | |
|-------------------------|----------|
| EZtime LAMP Premix (2X) | 0.625 ml |
| Fluorescence Dye (10X) | 0.125 ml |

Description

LAMP (Loop-mediated Isothermal Amplification) is a rapid, simple, economical, and highly sensitive nucleic acid amplification technique that can be performed under isothermal conditions (constant temperature), eliminating the need for expensive equipment such as thermal cyclers. Because it is very simple to operate and only requires a constant temperature instrument, so it is suitable for field screening, point-of-care or bedside testing, primary care institutions, small-scale clinical laboratories, remote areas with insufficient resources, field molecular epidemiological surveys or large-scale quarantine work.

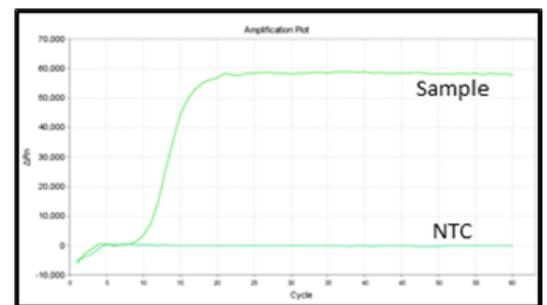
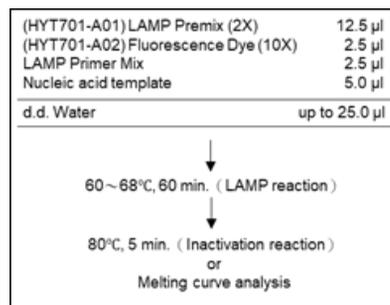
Features

- **Isothermal Reaction**---LAMP amplifies DNA at a constant temperature, removing the need for a thermal cycler.
- **Speed**---The amplification is rapid, often yielding results in 30-60 minutes, making it suitable for time-sensitive diagnostics.
- **High Specificity**---It uses 4 to 6 primers targeting 6 to 8 regions of the target DNA, ensuring high specificity.
- **High Sensitivity**---LAMP can detect even low amounts of DNA, making it effective for early-stage infection detection.

Applications

- **Clinical diagnostics**---Detection of infectious diseases such as COVID-19, tuberculosis, malaria, and other viral or bacterial pathogens.
- **Food safety and environmental monitoring**---Identification of foodborne pathogens and contamination in water or environmental samples.
- **Agriculture and veterinary medicine**---Detection of plant and animal pathogens for disease control.
- **Point-of-care testing**---Suitable for field or low-resource settings because it operates at a constant temperature and does not require complex equipment.

Results



The nucleic acid sample showed a signal reaction within 10 minutes; the negative control group showed no signal reaction within 60 minutes.

Related Products

- UniversAll™ Tissue Extraction/PCR Kits 1-1
- T&A Cloning Kit 3-14

Deoxynucleotides (dNTPs)

Description

Yeastern Biotech offers nucleotides with high purity for use in PCR, RT-PCR, RT assay, DNA labeling reactions and sequencing/cycle sequencing analysis.

Features

- Greater than 99% purity confirmed by HPLC.
- Free of trace contaminating nucleotides.
- Free of endo- and exodeoxyribonuclease, ribonuclease, phosphatase and nicking activities.
- Highly stable - the neutral pH of the nucleotide solutions ensures stability during long-term storage.
- Stable for years at -20°C.
- Stable after multiple freeze-thaw cycles.
- 90-95% of dNTPs remain in triphosphate form after 7 weeks at room temperature.
- 85-90% of dNTPs remain in triphosphate form after 30 cycles of PCR (1 min at 94°C; 3 mins at 72°C).
- Application tested in standard PCR, high fidelity PCR, long range PCR (40 kb), cDNA synthesis, RT-PCR, and real-time PCR.

Applications

Yeastern Biotech dNTPs can be used in all molecular biology applications including standard PCR, high fidelity, long PCR, LAMP-PCR, cDNA synthesis, RT-PCR, real-time PCR, RDA, MDA, DNA labeling and DNA sequencing.

Quality Control

- dCTP, dATP, dGTP and dTTP are all in the form of sodium salt (pH 8.3); > 99% dCTP (HPLC), < 0.9% dCDP.
- Greater than 99% purity of each component confirmed by HPLC. Functionally tested in PCR with Taq and Pfu DNA Polymerases. The absence of endo-, exodeoxyribonuclease, ribonuclease and nicking activities confirmed by appropriate tests.



-20°C

FYT013-200UL

dNTPs (10 mM) 200 µl

FYT014-100UL

dCTP (100 mM) 100 µl

FYT015-100UL

dATP (100 mM) 100 µl

FYT016-100UL

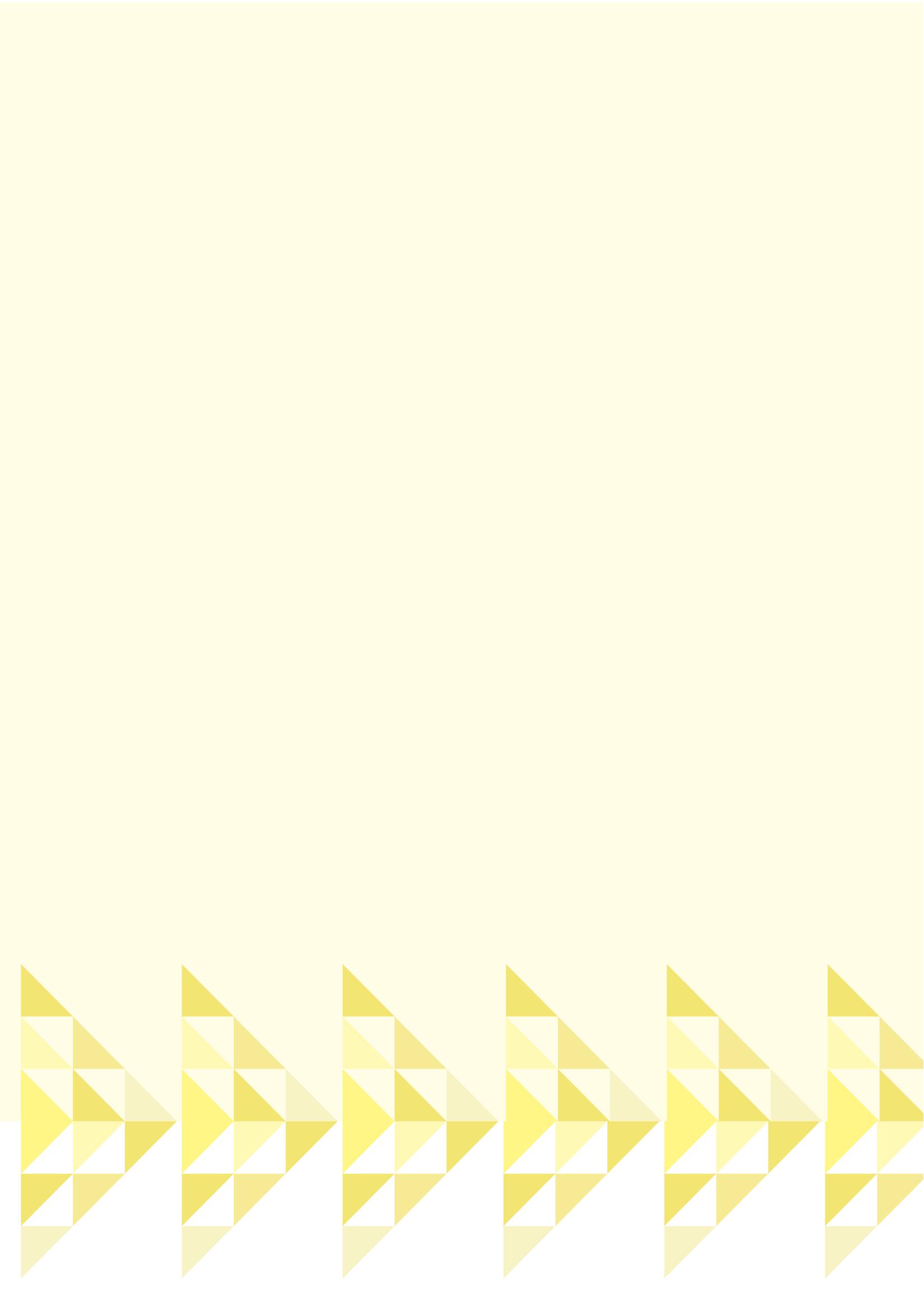
dGTP (100 mM) 100 µl

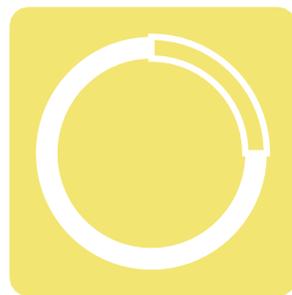
FYT017-100UL

dTTP (100 mM) 100 µl

Related Products

- UniversAll™ Tissue Extraction/PCR Kits 1-1
- YEAtaq DNA Polymerase 2-1
- T&A Cloning Kit 3-14





Gene Cloning

Introduction:

ECOS™ 1 Min Transformation Competent Cells

Description

ECOS™ Competent Cells are the first innovative products of Yeastern Biotech's R&D team, which strives to bring noble products that make research faster and easier. ECOS™ is Yeastern Biotech's registered trademark in USA, Japan, Canada, Korea, and Taiwan. The name speaks for the merits of this line of products - *E. Coli* **One-Step** transformation. The reverse of this name is SOCE - **SOC Eliminated**. ECOS™ protocol allows users to finish the transformation within one step and to skip the SOC recovery step. Therefore, ECOS™ competent cells are the fastest transforming cells worldwide. The traditional protocol requires recovery step and takes 1 ~ 2 hours to finish, but ECOS™ allows transformation to be finished within 1 minute and the procedures are a lot simpler. Please check the ECOS™ protocol section.

The latest progress in the ECOS™ product section is the development of another 3 protocols in addition to the standard ECOS™ protocol that was first introduced in 2003. The non-heat shock protocol allows transformation to be further simplified, and the 2 ~ 6-minute protocol enhances transformation efficiency significantly. These protocols have been developed based on the results of numerous experiments involving different antibiotics, plasmid size, and various conditions. Now, Yeastern Biotech offers our customers a new ECOS™ competent cells with similar transformation efficiency to that of electroporation for applications which require very high transformation efficiency such as the construction of genomic libraries.

ECOS™ Efficiency Guarantee

ECOS™ competent cells should be stored in a - 70°C freezer. Since competent cells are very sensitive to freeze-thaw cycles, exposure to temperature variations should be minimized. In order to control the quality of competent cells, Yeastern Biotech adds electronic temperature monitors to record the temperature during the shipping process. Yeastern Biotech is the only company that offers this service.

ECOS™ Technology is Patented

Yeastern Biotech Co., Ltd. owns the patent of ECOS™ technology (including protocol) exclusively in Canada (**TMA622,671**), USA (**US 6,864,088, US 7,098,033, US 7,820,443**), UK (**GB2383582**), German (**Nr. 102 51 429**), France (**FR 2832727**), Taiwan (**I 229696**), China (**ZL 2005 1 0112590.8**), Korea (**0604787, 10-1350283**), and Japan (**4867595**). Under the protection of this patent, Yeastern Biotech is the only company that has the right to use the protocol with the claims that transformation with our ECOS™ chemically competent cells can be finished within one minute and SOC is not needed. If any one intends to sell competent cells that use ECOS™ protocol or its similar version, please contact our headquarter in Taiwan for licensing.

Features

- No LB/SOC required
- Revolutionary 1- or 6-minute transformation, instead of 1 ~ 2 hours
- Procedures are simplified
- Several protocols to choose from according to your needs in efficiency and convenience
- Strict QC process to check the efficiency of each batch produced
- Ideal for bench users to automatize cloning projects
- Electronically monitored shipping process to check temperature fluctuation during shipping

Quality Control

Each lot of competent cells has to pass three quality control tests before shipping:

- 1. Efficiency test :** each batch of ECOS™ should meet the claimed transformation efficiency at the time of production using Protocol 1 or 3 (Page 3-2) and supercoiled pUC19 DNA.
- 2. Contamination test :** competent cells were plated directly on ampicillin plates without being transformed. To pass the test, no colonies should be seen after overnight incubation.
- 3. A-complementation test :** this test is performed for all ECOS™ competent cells except for ECOS™ 21(DE3) and ECOS™ 2163. To pass the test, the ratio of white colonies over the total colonies should be less than 3%.

GENE CLONING

Features

| Mutation | Applications | ECOS™ X / ECOS™ 101 [DH5α] | ECOS™ 9-5 [JM109] | ECOS™ Blue [XL1-Blue] | ECOS™ 21 [BL21(DE3)] | ECOS™ 10B [DH10B] | ECOS™ 2163 [GM2163] | ECOS™ St [HB101 Derived] | ECOS™ SONIC [BL21(DE3) Derived] |
|-------------|--|--|---|---|--|---|---|---|--|
| | | F ⁻ endA1 hsdR17 (rk ⁻ , mk ⁻) supE44 thi-1 λ ⁻ recA1 gyrA96 relA1 Δ(argF-lacZYA) U169 φ80d lacZ Δ M15 deoR | F ⁻ traD/6 proA+ proB+ lacIq Δ(lacZ) M15 Δ(lac - proAB) supE44 hsdR17 recA1 gyrA96 thi-1 endA1 relA1 e14 ⁻ λ ⁻ | F ⁻ recA1 endA1 gyrA96 thi-1 hsdR17(rk ⁻ , mk ⁻) supE44 λ ⁻ Δ(lac) proAB lacIqΔM15 Tn10 (tetr) | F ⁻ hsdS gal (λ clts857 ind1 Sam7 nin5 lacUV5- T7 gene I) | F ⁻ endA1 recA1 galE15 galK16 nupG rpsL ΔlacX74 φ80d lacZ ΔM15 araD1/9 Δ(lara,leu)7697 mcrA Δ(mrr hsdRMS ⁻ mcr BC) λ ⁻ | F ⁻ ara ⁻ 14 leuB6 thi ⁻ 1 fhuA/1 lacY1 tsx ⁻ 78 galK2 galT22 supE44 rpsL1/6(strr) xyl ⁻ 5 mtl ⁻ 1 dam1::Tn9 (camr) dcm ⁻ 6 mcrB1 hsdR2(rk ⁻ mk ⁻) mcrA | F ⁻ mcrB mrr hsdS20(rB ⁻ , mB ⁻) recA13 supE44 ara-14 galK2 lacY1 proA2 rpsL20(StrR) xyl-5 λ ⁻ leu mtl-1 | F ⁻ ompT hsdSB (rB ⁻ mB ⁻) gal dcm (DE3)ΔendA ΔrecA pLysS (CamR) |
| endA1 | Prevent DNA degradation during extraction | ✓ | ✓ | ✓ | ✗ | ✓ | ✗ | ✓ | ✓ |
| recA1 | Prevent DNA recombination | ✓ | ✓ | ✓ | ✗ | ✓ | ✗ | ✓ | ✓ |
| hsdR | Improve transformation efficiency (for unmethylated PCR DNA and cDNA) | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ |
| deoR | Improve transformation efficiency (for large size plasmid and cosmids) | ✓ | ✗ | ✗ | ✗ | ✓ | ✗ | ✗ | ✗ |
| LacZ M15 | Inhibit LacZ gene expression for blue/white screen | ✓ | ✓ | ✓ | ✗ | ✓ | ✗ | ✓ | ✗ |
| rne131 | Inhibit RNase E and improve mRNA stability | ✗ | ✗ | ✗ | ✓ | ✗ | ✗ | ✗ | ✓ |
| ompT & Lon | ompT & Lon Protease deficient and improve protein yield | ✗ | ✗ | ✗ | ✓ | ✗ | ✗ | ✗ | ✓ |
| dam / dcm | Prevent DNA methylation | ✗ | ✗ | ✗ | ✓ / ✗ | ✗ | ✓ / ✓ | ✗ | ✓ / ✗ |
| mcrA / mcrB | Prevent methylated DNA from degradation | ✗ | ✗ | ✗ | ✗ | ✓ / ✓ | ✓ / ✓ | ✗ / ✓ | ✗ |

ECOS™ Protocol Comparison Table

| Cloning Applications | ECOS™ X / ECOS™ 101 [DH5α] | ECOS™ 9-5 [JM109] | ECOS™ Blue [XL1-Blue] | ECOS™ 21 [BL21(DE3)] | ECOS™ 10B [DH10B] | ECOS™ 2163 [GM2163] | ECOS™ St [HB101 Derived] | ECOS™ SONIC [BL21(DE3) Derived] |
|---------------------------|----------------------------|-------------------|-----------------------|----------------------|-------------------|---------------------|--------------------------|---------------------------------|
| Large Plasmids > 6 kb | Ideal | * | * | * | Yes | Yes | Yes | * |
| Subcloning | Ideal | Yes | Yes | * | Ideal | No | Yes | Yes |
| cDNA Library | Yes | Yes | Yes | * | Yes | * | Yes | * |
| Fast Growth | * | Ideal | * | Yes | * | * | * | Yes |
| Single Stranded DNA | * | Ideal | * | * | * | * | * | * |
| Mutagenesis | Yes | * | * | No | Yes | * | Yes | * |
| Protein Expression | No | No | No | Ideal | * | No | No | Yes |
| Toxic Protein Expression | No | No | No | No | No | No | No | Ideal |
| Blue/White Screen | Yes | Yes | Ideal | No | Yes | No | Yes | No |
| DNA Unmethylation | No | No | No | No | No | Yes | No | No |
| Genomic DNA Cloning | No | No | No | No | Yes | No | No | No |
| Unstable Sequence Cloning | No | No | No | No | No | No | Ideal | No |

* Means the strain can be used for the purpose but may not yield the best result.

ECOS™ X Competent Cells

Strain: DH5α



-70°C

FYE610-10VL (10 preps)

Efficiency >5 x 10⁹ cfu/μg

100 μl/vial

10 vials

Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE610-80VL (80 preps)

Efficiency >5 x 10⁹ cfu/μg

100 μl/vial

80 vials

Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

Description

DH5α is the most popular *E. coli* strain for everyday cloning applications. It supports blue/white screening for easy selection of recombinant DNA with X-Gal. In addition, DH5α carries *recA1* mutation that eliminates homologous recombination ensuring insert stability. It also carries *endA1* mutations that greatly improve the quality of plasmid DNA and yield prepared from mini-prep. It is useful for the transformation of large plasmids and two-hybrid systems (up to 20 kb).

After 5-year invention based on our ECOS™ technology, Yeastern Biotech is able to offer our customers a brand new competent cell product with similar transformation efficiency to that of electroporation. ECOS™ X can be utilized for applications which require very high transformation efficiency such as the construction of genomic libraries. The “X” as shown in the name, stands for extreme, meaning this competent cell was designed to exceed limitation and to set new limitation! The “X” also represents “10” in Roman numeral; our new product aims to reach 10¹⁰ in its efficiency.

Genotype

F⁻ *endA1 hsdR17*(rk⁻, mk⁻) *supE44 thi-1 λ⁻ recA1 gyrA96 relA1 Δ(argF-lacZYA)*
U169 Φ80d *lacZ ΔM15 deoR*

Features

- One step in one tube and finish in 6 minutes
- High efficiency as electro-competent cells
- Requires no expensive equipment

Applications

- Efficient transformation of products from routine TA cloning
- Transformation of minute amounts of PCR products
- Construction of a more representative gene library

Efficiency

> 5 x 10⁹ cfu/μg

Recommended for:

- Super high efficiency cloning
- Cloning lowest abundance cDNA
- Gene library

Related Products

| | |
|--------------------------------|------|
| • HiYield Plasmid Mini Kit 2.0 | 1-4 |
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • Glass Plating Beads | 3-26 |

Do NOT store the cells in liquid nitrogen!

ECOS™ 101 Competent Cells

Strain: DH5α

Description

DH5α is the most popular *E. coli* strain for everyday cloning applications. It supports blue/white screening for easy selection of recombinant DNA with X-Gal. In addition, DH5α carries *recA1* mutation that eliminates homologous recombination ensuring insert stability. It also carries *endA1* mutations that greatly improve the quality of plasmid DNA and yield prepared from mini-prep. It is useful for the transformation of large plasmids and two-hybrid systems (up to 14 kb).

Genotype

F^- *endA1 hsdR17*(rk^- , mk^-) *supE44 thi-1 λ^-recA1 gyrA96 relA1 Δ*(*argF-lacZYA*) U169 Φ80d *lacZ ΔM15 deoR*

Features

Suitable for cloning with large plasmid and cDNA library construction, and also allow blue-white colony screening.

Applications

- Cloning and subcloning
- Scale-up application
- Blue/white screening

Efficiency

> 1 x 10⁸ cfu/μg

Recommended for:

- Subcloning
- General cloning

>3 x 10⁸ cfu/μg

Recommended for:

- High efficiency cloning
- Cloning low abundance transcripts
- General cloning
- Gene library

> 1 x 10⁹ cfu/μg

Recommended for:

- Super high efficiency cloning
- Cloning lowest abundance transcripts
- Gene library

Do NOT store the cells in liquid nitrogen!



-70°C

FYE607-10VL (10 preps)

Efficiency >1 x 10⁸ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE607-80VL (80 preps)

Efficiency >1 x 10⁸ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE608-10VL (10 preps)

Efficiency >3 x 10⁸ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE608-80VL (80 preps)

Efficiency >3 x 10⁸ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE609-10VL (10 preps)

Efficiency >1 x 10⁹ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE609-80VL (80 preps)

Efficiency >1 x 10⁹ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE607-96WL (96 preps)

Efficiency >5 x 10⁷ cfu/μg

50 μl/well 96 wells

FYE608-96WL (96 preps)

Efficiency >1.5 x 10⁸ cfu/μg

50 μl/well 96 wells

FYE609-96WL (96 preps)

Efficiency >5 x 10⁸ cfu/μg

50 μl/vial 96 wells

FYE678-10VL (10 preps)

Efficiency >5 x 10⁸ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE678-80VL (80 preps)

Efficiency >5 x 10⁸ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

Related Products please refer to 3-5

ECOS™ 9-5 Competent Cells

Strain: JM109



-70°C

FYE707-10VL (10 preps)

Efficiency >5 x 10⁷ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE707-80VL (80 preps)

Efficiency >5 x 10⁷ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE708-10VL (10 preps)

Efficiency >1 x 10⁸ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE708-80VL (80 preps)

Efficiency >1 x 10⁸ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE709-10VL (10 preps)

Efficiency >5 x 10⁸ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE709-80VL (80 preps)

Efficiency >5 x 10⁸ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE707-96WL (96 preps)

Efficiency >2.5 x 10⁷ cfu/μg

50 μl/well 96 wells

FYE708-96WL (96 preps)

Efficiency >5 x 10⁷ cfu/μg

50 μl/well 96 wells

FYE709-96WL (96 preps)

Efficiency >2.5 x 10⁸ cfu/μg

50 μl/vial 96 wells

Related Products

| | |
|--------------------------------|------|
| • HiYield Plasmid Mini Kit 2.0 | 1-4 |
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • Glass Plating Beads | 3-26 |

Description

JM109, a K strain bacterium, carries mutations in *recA* and *endA*, leading to minimal recombination and improved quality of isolated plasmid DNA. In addition, the cells carry the F' episome which allows blue/white screening for recombinant DNA. Thus, it has been a popular strain for routine subcloning. The cells can also be utilized for single-stranded DNA rescue when M13 or phagemid systems are used simultaneously. The JM109 strain is sensitive to all common antibiotics.

This product can become a powerful cloning tool when it is used in combination with Yeastern's rapid ligation kit (**#FYC003-100R**). The ligation can be finished within 5 minutes and followed by 1-minute transformation. The fast growth rate of JM109 allows colonies to show up within 8 hours. Try this strain when you are in urgent.

Genotype

F' *traD16 proA⁺ proB⁺ lacIq Δ(lacZ)M15 Δ(lac-proAB) supE44 hsdR17 recA1 gyrA96 thi⁻1 endA1 relA1 e14⁻ λ⁻*

Features

- Rapid growing strain
- Carries F' episome

Applications

- Preparation of ssDNA
- Construction of gene library
- Subcloning

Efficiency

> **1 x 10⁸ cfu/μg**

Recommended for:

- High efficiency cloning
- Cloning low abundance transcripts
- General cloning
- Gene library

> **1 x 10⁹ cfu/μg**

Recommended for:

- High efficiency cloning
- Cloning low abundance transcripts
- General cloning
- Gene library

> **5 x 10⁷ cfu/μg**

Recommended for:

- Subcloning
- General cloning

Do not store the cells in liquid nitrogen!

ECOS™ Blue Competent Cells

Strain: XL1-Blue

Description

XL1-Blue is the most popular strain for blue/white screening. It is also an excellent host strain for routine cloning application using plasmid or lambda vectors. XL1-Blue cells are endonuclease (*endA*) deficient, which greatly improve the quality of mini-prep DNA, and are recombination (*recA*) deficient, improving insert stability. The *hsdR* mutation prevents the cleavage of cloned DNA by the *EcoK* endonuclease system. The *lacIqZΔM15* gene on the F' episome allows blue-white color screening.

Genotype

F' *recA1 endA1 gyrA96 thi⁻¹ hsdR17(rk⁻, mk⁺) supE44 λ⁻ Δ(lac) proAB lacIqZΔM15 Tn10 (tet')*

Features

- A strain modified to be ideal for blue/white screening
- Tetracycline resistant

Applications

- Routine cloning application
- Subcloning
- cDNA library
- Blue/white screening

Efficiency

> **5 x 10⁷ cfu/μg**

Recommended for:

- Routine cloning and subcloning
- Blue/white screening

> **2 x 10⁹ cfu/μg**

Recommended for:

- Routine cloning and subcloning
- Blue/white screening

> **5 x 10⁸ cfu/μg**

Recommended for:

- Routine cloning and subcloning
- Blue/white screening
- cDNA library



-70°C

FYE107-10VL (10 preps)

Efficiency >1 x 10⁸ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE107-80VL (80 preps)

Efficiency >1 x 10⁸ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE108-10VL (10 preps)

Efficiency >5 x 10⁸ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE108-80VL (80 preps)

Efficiency >5 x 10⁸ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE109-10VL (10 preps)

Efficiency >2 x 10⁹ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE109-80VL (80 preps)

Efficiency >2 x 10⁹ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE107-96WL (96 preps)

Efficiency >5 x 10⁷ cfu/μg

50 μl/well 96 wells

FYE108-96WL (96 preps)

Efficiency >2.5x 10⁸ cfu/μg

50 μl/well 96 wells

FYE109-96WL (96preps)

Efficiency >1 x 10⁹ cfu/μg

50 μl/vial 96 wells

Related Products

- | | |
|--------------------------------|------|
| • HiYield Plasmid Mini Kit 2.0 | 1-4 |
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • Glass Plating Beads | 3-26 |

Do NOT store the cells in liquid nitrogen!

ECOS™ 21 Competent Cells

Strain: BL21(DE3)



-70°C

FYE207-5VL (5 preps)

Efficiency > 2 x 10⁷ cfu/μg

100 μl/vial 5 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE207-40VL (40 preps)

Efficiency > 2 x 10⁷ cfu/μg

100 μl/vial 40 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE207-96WL (96 preps)

Efficiency > 1 x 10⁷ cfu/μg

50 μl/well 96 wells

Description

This strain provides high levels of protein expression. This strain carries the lambda DE3 lysogen, which expresses T7 RNA polymerase from the *lacUV5* promoter by isopropyl-1-thio-β-D-galactopyranoside (IPTG) induction. The mutated *rne* gene (*rne131*) encodes a truncated RNase E enzyme that lacks the ability to degrade mRNA, resulting in an increase in mRNA stability. The transformation efficiency of BL21 is usually low, so the 6 minutes/ heat shock cold plating protocol is recommended if high efficiency is desired.

Genotype

F⁻ *hsdS gal (λ clts857 ind1 Sam7 nin5 lacUV5-T7 gene l)*

Features

- Lack of Lon and Omp proteases allows high-level protein expression and accumulation
- Easy gene induction

Applications

- IPTG induced gene expression in *E. coli*
- Ideal for high level expression of non-toxic recombinant protein expression using T7-based expression vectors
- Subcloning

Efficiency

> 2 x 10⁷ cfu/μg

Recommended for:

- Recombinant protein expression
- Subcloning

Related Products

| | |
|--------------------------|------|
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • T&A™ Expression Vector | 3-17 |
| • Glass Plating Beads | 3-26 |

Do NOT store the cells in liquid nitrogen!

ECOS™ 10B Competent Cells

Strain: DH10B

Description

DH10B is an MC1061 derivative. This strain was designed for the propagation of large insert DNA library clones. It is used extensively, taking advantage of properties such as high DNA transformation efficiency and maintenance of large plasmids.

While DH10B has been classically reported to be *galU galK*, the preliminary genome sequence for DH10B indicates that DH10B is actually *galE galK galU+*. Genome sequence indicates that DH10B is actually *deoR+*. Complete genome sequence has been published.

GenoType

F^- *endA1 recA1 galE15 galK16 nupG rpsL ΔlacX74* Φ 80d *lacZ ΔM15 araD1/9 Δ(lara,leu)7697 mcrA Δ(mrr⁻hsdRMS⁻mcrBC) λ⁻*

FeaTures

- Streptomycin resistant
- High transformation efficiency

ApplicaTions

- Suitable for genomic DNA cloning
- Allow large inserts
- Cloning of methylated cytosine or adenine containing DNA
- Blue/white screening

Efficiency

> **5×10^7 cfu/μg**

Recommended for:

- Subcloning
- Genomic DNA cloning

> **2×10^8 cfu/μg**

Recommended for:

- Cloning of large inserts
- Genomic DNA cloning



-70°C

FYE507-10VL (10 preps)

Efficiency > 5×10^7 cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10^{-4} μg/μl)

FYE507-80VL (80 preps)

Efficiency > 5×10^7 cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10^{-4} μg/μl)

FYE508-10VL (10 preps)

Efficiency > 1×10^8 cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10^{-4} μg/μl)

FYE508-80VL (80 preps)

Efficiency > 1×10^8 cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10^{-4} μg/μl)

FYE507-96WL (96 preps)

Efficiency > 2.5×10^7 cfu/μg

50 μl/well 96 wells

Do NOT store the cells in liquid nitrogen!

Related Products

- | | |
|--------------------------------|------|
| • HiYield Plasmid Mini Kit 2.0 | 1-4 |
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • Glass Plating Beads | 3-26 |

ECOS™ 2163 Competent Cells

Strain: GM2163



FYE807-10VL (10 preps)

Efficiency > 1 x 10⁷ cfu/μg

100 μl/vial 10 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE807-80VL (80 preps)

Efficiency > 1 x 10⁷ cfu/μg

100 μl/vial 80 vials
Control Plasmid (pUC19) 5 μl (10⁻⁴ μg/μl)

FYE807-96WL (96 preps)

Efficiency > 5 x 10⁶ cfu/μg

50 μl/well 96 wells

Description

GM2163 is an *E. coli* K12 strain, which is deficient in both *dam* and *dcm* genes thus is suitable for the propagation of plasmid or cloned DNA to be cut with Dam or Dcm-sensitive restriction enzymes. This strain is not recommended as a host for primary cloning/ligation since the *dam* mutation can result in higher mutation rates and a reduction in the transformation efficiency.

Genotype

F⁻ *ara*⁻14 *leu*B6 *thi*⁻1 *fhu*A/1 *lac*Y1 *tsx*⁻78 *gal*K2 *gal*T22 *sup*E44 *rps*L1/6(*str*r) *xy*L⁻5 *mt*L⁻1 *dam*1/:Tn9 (*cam*r) *dcm*⁻6 *mcr*B1 *hsd*R2(*rk*⁻*mk*⁺) *mcr*A

Features

- Chloramphenicol resistant
- Not suitable for blue/white screening
- Not recommended as a host for primary cloning/ligation

Applications

- Propagation of plasmid free of Dam and Dcm methylations

Efficiency

> **1 x 10⁷ cfu/μg**

Recommended for:

- Propagation of plasmid to be cut with Dam or Dcm-sensitive restriction enzymes

Related Products

| | |
|--------------------------------|------|
| • HiYield Plasmid Mini Kit 2.0 | 1-4 |
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • Glass Plating Beads | 3-26 |

Do NOT store the cells in liquid nitrogen!

ECOS™ St

Strain: HB101 Derived

Description

ECOS™ St competent cells is a strain derived from HB101 and suitable for cloning unstable inserts such as lentiviral DNA with LTR (Long terminal repeat). There are two transformation efficiencies of this strain in ECOS™ product line. If you want to transform longer plasmids, plasmid with a kanamycin selection system, or get maximum quantity colonies, you can choose HYE007-S08 which the efficiency is 1×10^9 cfu/ μ g.

Genotype

F^- mcrB mrr hsdS20(rB^- , mB^-) recA13 supE44 ara-14 galK2 lacY1 proA2 rpsL20(Str^R) xyl-5 λ^- leu mtl-1

Features

ECOS™ St carry the recA13 mutation, which lowers nonspecific recombination and helps maintain unstable inserts like lentiviral vectors with direct repeats.

Applications

- Lentiviral/Retroviral construct
- Unstable sequence cloning
- Direct replacement for Invitrogen Stbl3™ and Lucigen Endura™

*The trademarks belong to Thermo Fisher and Lucigen.

Efficiency

> 1×10^8 cfu/ μ g

Recommended for:

- Cloning unstable inserts
- General cloning

> 1×10^9 cfu/ μ g

Recommended for:

- High efficiency cloning
- Cloning unstable inserts
- General cloning



-70°C

FYE307-10VL (10 preps)

Efficiency > 1×10^8 cfu/ μ g

100 μ l/vial 10 vials
Control Plasmid (pUC19) 5 μ l (10^{-4} μ g/ μ l)

FYE307-80VL (80 preps)

Efficiency > 1×10^8 cfu/ μ g

100 μ l/vial 80 vials
Control Plasmid (pUC19) 5 μ l (10^{-4} μ g/ μ l)

FYE309-10VL (10 preps)

Efficiency > 1×10^9 cfu/ μ g

100 μ l/vial 10 vials
Control Plasmid (pUC19) 5 μ l (10^{-4} μ g/ μ l)

FYE309-80VL (80 preps)

Efficiency > 1×10^9 cfu/ μ g

100 μ l/vial 80 vials
Control Plasmid (pUC19) 5 μ l (10^{-4} μ g/ μ l)

Related Products

- | | |
|--------------------------------|------|
| • HiYield Plasmid Mini Kit 2.0 | 1-4 |
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • Glass Plating Beads | 3-26 |

Do NOT store the cells in liquid nitrogen!

ECOS™ SONIC Competent Cells

Strain:BL21(DE3) Derived



FYE907-5VL (5 preps)

Efficiency > 1×10^8 cfu/ μ g

100 μ l/vial 5 vials
Control Plasmid (pUC19) 5 μ l (10^{-4} μ g/ μ l)

FYE907-20VL (20 preps)

Efficiency > 1×10^8 cfu/ μ g

100 μ l/vial 20 vials
Control Plasmid (pUC19) 5 μ l (10^{-4} μ g/ μ l)

Description

ECOS™ Sonic pLysS competent cells is the derivative of BL21(DE3) pLysS strain. It inherits the protein expression, and the fast-growing feature from its parent strain and pLysS strain can express T7 lysozyme to reduce the activity of T7 RNA polymerase that allows the host including toxic gene under the control of T7 promoter not to be expressed until induction. The further deletion of endA and recA improve the quality of plasmid DNA and enable the plasmid extraction in 3-6 hours growing.

T&A Expression Kit (FYC201-10P) is recommended for ECOS™ Sonic; it combines the primary cloning of PCR product, and the subcloning for protein expression make the experiment easier and faster.

Genotype

F⁻ ompT hsdS_B (r_B⁻ m_B⁻) gal dcm (DE3) Δ endA Δ recA pLysS (Cam^R)

Features

- Protein expression and cloning in one strain
- Reduced recombination and DNA degradation
- Protease deficiency for better protein yields

Applications

- T7-driven protein expression
- Toxic or tightly regulated gene expression
- Clone → express without strain switching

Efficiency

> 1×10^8 cfu/ μ g

Recommended for:

- Recombinant protein expression
- Subcloning

Related Products

| | |
|--------------------------------|------|
| • HiYield Plasmid Mini Kit 2.0 | 1-4 |
| • YEAtaq DNA Polymerase | 2-1 |
| • T&A™ Cloning Kit | 3-14 |
| • T&ATM Expression Vector | 3-17 |
| • Glass Plating Beads | 3-26 |

Do NOT store the cells in liquid nitrogen!

T&A™ Cloning Kit

T&A™ Cloning Kit II

Description

Molecular cloning assisted by vectors is the most popular and common method to obtain genes of interest. Yeastern Biotech's T&A™ Cloning Kit offers a quick, reliable and efficient method for cloning a variety of DNA sequences.

The T&A™ Cloning Kit (**FYC001-20P**) contains the T&A™ Cloning Vector and all the reagents needed for ligation. It is a convenient pack for cloning PCR product generated using thermostable DNA polymerases, such as YEAtaq DNA polymerase, which add a single terminal 3'-dA nucleotide overhang. After ligation, the mixture can be used directly for transformation into competent cells (ECOS™) or be purified first to achieve higher transformation efficiency.

Recently, YB has designed a new cloning vector for user convenience. The new T&A™ Cloning Vector II (**FYC101-20P**) consist of 2 *EcoR* I cutting sites (441, 500) within the multiple cloning site.

Features

- Fast ligation, completed in only 5 minutes
- High transformation efficiency
- More accurate results
- Accept a wide range of inserts with different sizes
- Two types of ligation buffers provided for your convenience
- Allow blue/white screening
- Contain ampicillin marker for antibiotic selection
- Include M13 primer sites for convenient sequencing

Applications

Cloning of terminal 3'-dA nucleotides overhang PCR products up to 5 kb

Quality Control

- DNA concentration of the vectors is 25 ng/μl
- The absorbance ratio (A_{260}/A_{280}) is between 1.6-2.0
- The size of the vectors is about 2.7 kb
- The colony number of background control is less than 50 when the transformation efficiency of competent cells is 1×10^8 cfu/μg DNA
- The colony number ratio of self-ligation control to positive control is less than 15%
- The colony number of positive control is more than 500 when the transformation efficiency of competent cells is 5×10^8 cfu/μg DNA
- The ligation correctness with the control insert into the vectors is more than 87.5%



-20°C

FYC001-20P (20 preps)

T&A™ Cloning Kit

| | |
|------------------------|------------------|
| T&A™ Cloning Vector | 40 μl (25 ng/μl) |
| Control Insert DNA | 10 μl (10 ng/μl) |
| yT4 DNA Ligase | 20 μl |
| 10× Ligation Buffer A | 50 μl |
| 10× Ligation Buffer B | 50 μl |
| Forward Primer (M13-F) | 50 μl (10 μM) |
| Reverse Primer (M13-R) | 50 μl (10 μM) |

FYC002-20P (20 preps)

| | |
|---------------------|------------------|
| T&A™ Cloning Vector | 40 μl (25 ng/μl) |
| Control Insert DNA | 10 μl (10 ng/μl) |

FYC101-20P (20 preps)

T&A™ Cloning Kit II

| | |
|------------------------|------------------|
| T&A™ Cloning Vector II | 40 μl (25 ng/μl) |
| Control Insert DNA | 10 μl (10 ng/μl) |
| yT4 DNA Ligase | 20 μl (2 U/μl) |
| 10× Ligation Buffer A | 50 μl |
| 10× Ligation Buffer B | 50 μl |
| Forward Primer (M13-F) | 50 μl (10 μM) |
| Reverse Primer (M13-R) | 50 μl (10 μM) |

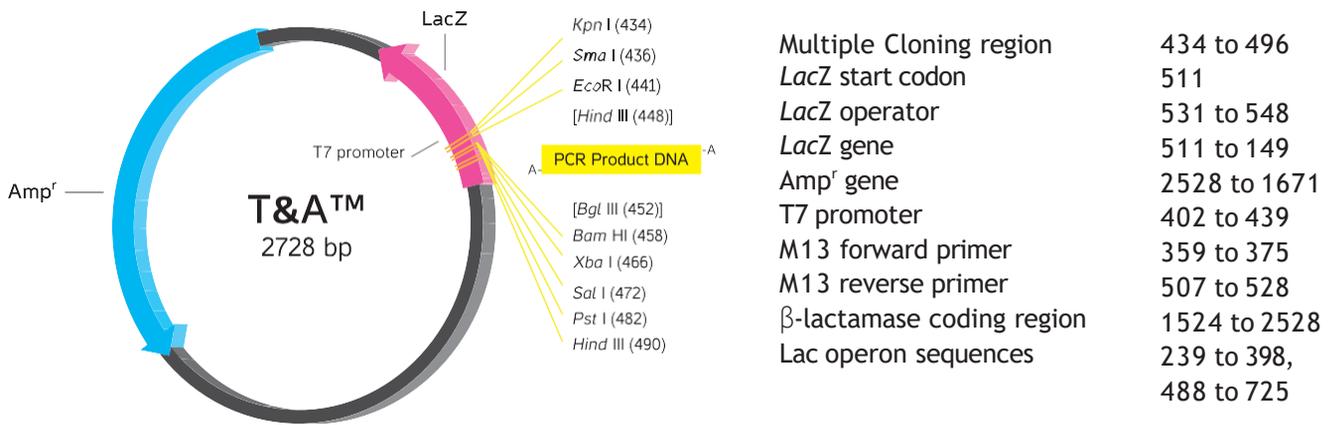
FYC102-20P (20 preps)

| | |
|------------------------|------------------|
| T&A™ Cloning Vector II | 40 μl (25 ng/μl) |
| Control Insert DNA | 10 μl (10 ng/μl) |

Related Products

| | |
|---------------------------------------|------|
| • YEA Ladder DNA Markers | 1-12 |
| • YEAtaq DNA Polymerase | 2-1 |
| • Deoxy+ HiSpec Reverse Transcriptase | 2-5 |
| • Deoxynucleotides (dNTPs) | 2-10 |
| • ECOS™ Competent cells | 3-1 |

Map and Sequence reference points of the T&A™ Cloning Vector



* Before the insert incorporate into the T&A™ Cloning Vector, there is only one *Hind* III site and no *Bgl* II site. After the incorporation, the T and A nucleotide on the insert will complement the sequence on the vector and generate these two new sites. This merit of T&A™ Cloning Vector makes cloning more economic and convenient.

```

301 TACGCCAGCT GCGCAAAGGG GGATGTGCTG CAAGGCGATT AAGTTGGGTA
    ATGCGGTGCA CCGCTTTCCC CCTACACGAC GTTCCGCTAA TTCAACCCAT

          M13 Forward Primer
351 ACGCCAGGGT TTCCCAGTC ACGACGTTGT AAAACGACGG CCAGTGAATT
    TCGGTTCCCA AAAGGGTCAG TGCTGCAACA TTTTGCTGCC GGTCACCTAA

          T7 Promoter
401 GTAATACGAC TCACTATAGG GCGAGCTCGG TACCCGGGCG AATTCCAAGC
    CATTATGCTG AGTGATATCC CGCTCGAGCC ATGGGCCCGC TTAAGGTTCG

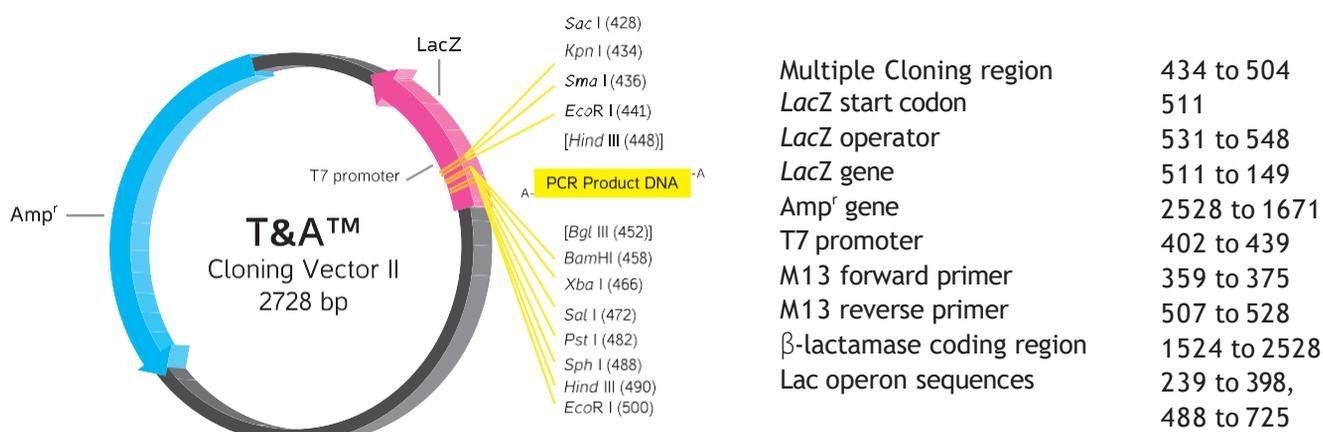
          Bgl II BamHI Xba I Sal I Pst I
451 T T Insert AGATCTGGAT CCCCTCTAGA GTCGACCTGC AGGCATGCAA
    A A DNA TCTAGACCTA GGGGAGATCT CAGCTGGACG TCCGTACGTT

          Hind III
493 GCTTGGCGTA ATCATGGTCA TAGCTGTTTC CTGTGTGAAA TTGTTATCCG
    CGAACCCGAT TAGTACCAGT ATCGACAAAG GACACACTTT AACAATAGGC

          M13 Reverse Primer
    
```

| Enzyme | Position | Enzyme | Position | Enzyme | Position | Enzyme | Position | Enzyme | Position |
|---------|----------|--------|----------|-----------|----------|--------|----------|--------|----------|
| Aat II | 2664 | AspE I | 1742 | Cfr10 I | 1822 | Mam I | 457 | Ssp I | 2546 |
| Acc65 I | 430 | Ava I | 434 | Dra II | 2718 | Nar I | 237 | Xba I | 466 |
| Acc I | 473 | Ban II | 428 | Eam1105 I | 1742 | Nde I | 185 | Xma I | 434 |
| Acs I | 441 | BamH I | 458 | Ecl136 II | 426 | Pst I | 482 | Xmn I | 2341 |
| Afl III | 849 | Bcg I | 2281 | Eco0109 I | 2718 | Sac I | 428 | | |
| Ahd I | 1742 | Bpm I | 1812 | EcoR I | 441 | Sal I | 472 | | |
| AlwN I | 1265 | BsaB I | 457 | Hinc II | 474 | Sap I | 733 | | |
| Apo I | 441 | Bsa I | 1803 | Hind II | 474 | Sca I | 2222 | | |
| Asp700 | 2341 | BspM I | 485 | Kas I | 236 | Sma I | 436 | | |
| Asp718 | 430 | BsrF I | 1822 | Kpn I | 434 | Sph I | 488 | | |

Map and Sequence reference points of the T&A™ Cloning Vector II



* Before the insert incorporate into the T&A™ Cloning Vector II, there is only one *Hind* III site and no *Bgl* II site. After the incorporation, the T and A nucleotide on the insert will complement the sequence on the vector and generate these two new sites. This merit of T&A™ Cloning Vector II makes cloning more economic and convenient.

| | | | | | | |
|-----|------------|------------|-------------|------------|------------|------------|
| 301 | TACGCCAGCT | GCGCAAAGGG | GGATGTGCTG | CAAGGCGATT | AAGTTGGGTA | |
| | ATGCGGTCSA | CCGCTTTCCC | CCTACACGAC | GTTCCGCTAA | TTCAACCCAT | |
| | | | | | | |
| | | | | | | |
| 351 | ACGCCAGGGT | TTCCTCAGTC | ACGACGTTGT | AAAACGACGG | CCAGTGAATT | |
| | TGCGGTCCCA | AAAGGGTCAG | TGCTGCAACA | TTTTGCTGCC | GGTCACTTAA | |
| | | | | | | |
| | | | | | | |
| 401 | GTAATACGAC | TCACTATAGG | GCGAGCTCGG | TACCCGGGCG | AATTCCAAGC | |
| | CATTATGCTG | AGTGATATCC | CGCTCGAGCC | ATGGGCCCGC | TTAAGGTTCG | |
| | | | | | | |
| | | | | | | |
| 451 | T T | Insert DNA | A GATCTGGAT | CCCTCTAGA | GTCGACCTGC | AGGCATGCAA |
| | A A- | | T CTAGACCTA | GGGGAGATCT | CAGCTGGACG | TCCGTACGTT |
| | | | | | | |
| | | | | | | |
| 491 | GCTTGGCGGA | ATTCTGGTCA | TAGCTGTTTC | CTGTGTGAAA | TTGTTATCCG | |
| | CGAACCCGCT | TAAGACCAGT | ATCGACAAAG | GACACACTTT | AACAATAGGC | |

| Enzyme | Position | Enzyme | Position | Enzyme | Position | Enzyme | Position | Enzyme | Position |
|-----------------|----------|----------------|----------|-------------------|----------|--------------|----------|--------------|----------|
| <i>Aat</i> II | 2664 | <i>Ava</i> I | 434 | <i>Bsr</i> F I | 1822 | <i>Kas</i> I | 236 | <i>Sap</i> I | 733 |
| <i>Acc</i> 65 I | 430 | <i>Bam</i> H I | 458 | <i>Cfr</i> 10 I | 1822 | <i>Kpn</i> I | 434 | <i>Sca</i> I | 2222 |
| <i>Acc</i> I | 473 | <i>Ban</i> II | 428 | <i>Dra</i> II | 2718 | <i>Mam</i> I | 457 | <i>Sma</i> I | 436 |
| <i>Afl</i> III | 849 | <i>Bcg</i> I | 2281 | <i>Eam</i> 1105 I | 1742 | <i>Nar</i> I | 237 | <i>Sph</i> I | 488 |
| <i>Ahd</i> I | 1742 | <i>Bpm</i> I | 1812 | <i>Ecl</i> 136 II | 426 | <i>Nde</i> I | 185 | <i>Ssp</i> I | 2546 |
| <i>Alw</i> N I | 1265 | <i>Bsa</i> B I | 457 | <i>Eco</i> 0109 I | 2718 | <i>Pst</i> I | 482 | <i>Xba</i> I | 466 |
| <i>Asp</i> 700 | 2341 | <i>Bsa</i> I | 1803 | <i>Hinc</i> II | 474 | <i>Sac</i> I | 428 | <i>Xma</i> I | 434 |
| <i>Asp</i> 718 | 430 | <i>Bsp</i> M I | 485 | <i>Hind</i> II | 474 | <i>Sal</i> I | 472 | <i>Xmn</i> I | 2341 |
| <i>Asp</i> E I | 1742 | | | | | | | | |

T&A™ Expression Vector



FYC211-10P (10 preps)

T&A™ Expression Vector 20 µl (25 ng/µl)

Control Insert DNA 10 µl (10 ng/µl)

Description

Molecular cloning assisted by vectors is the most popular and common method to obtain genes of interest. Yeastern Biotech's T&A™ Expression Vector offers a quick, reliable and efficient method for cloning a variety of DNA sequences.

The T&A™ Expression Vector (FYC211-10P) designed for direct cloning of PCR-amplified inserts (Taq polymerase products with 3' adenine (A) overhangs.) The vector contains complementary 3' thymine (T) overhangs to facilitate ligation without the need for restriction enzyme digestion. This vector drives recombinant protein expression under control of T7 promoter in *Escherichia coli*. Includes optional fusion 6×His tag sequences to enable affinity purification of the expressed protein. After ligation, the mixture can be used directly for transformation into competent cells (ECOS™ 21 or ECOS™ SONIC) to protein expression.

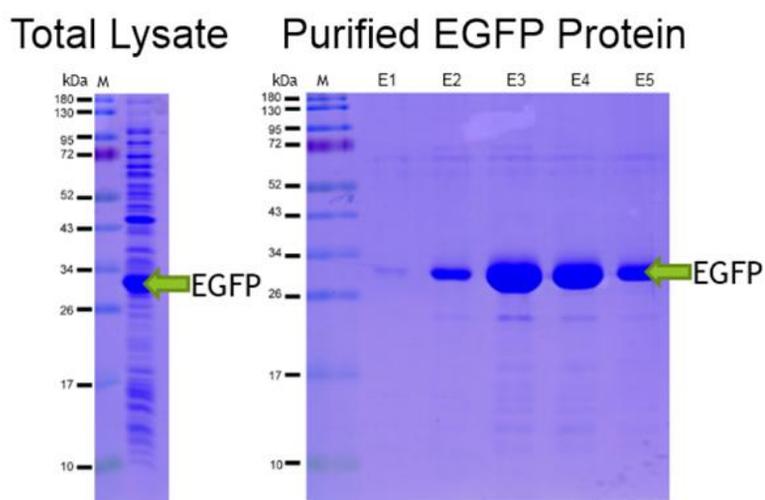
Features

- Able to express protein after TA Cloning
- His-tagged for purifying protein
- T7 promoter for highly expressing protein.
- Ampicillin marker for antibiotic and for fast operation of ECOS

Applications

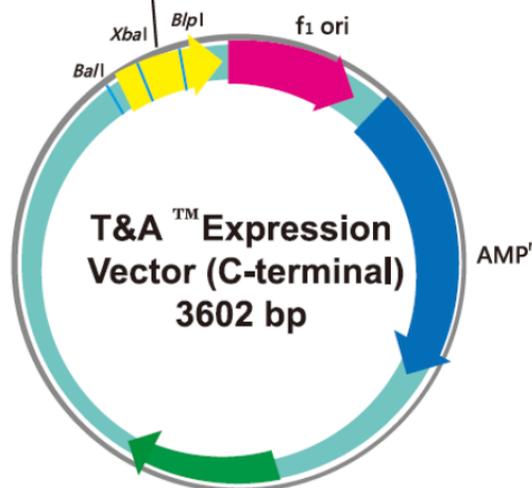
- Recombinant Protein Production
- Rapid Screening of PCR Products

Result



Related Products

- YEAtaq DNA Polymerase 2-1
- Deoxy+ HiSpec Reverse Transcriptase 2-5
- Deoxynucleotides (dNTPs) 2-10
- ECOS™ Competent cells 3-1



| | |
|-----------------------|-------------|
| ATG start codon | 3428 |
| T7 promoter | 3348 ~ 3366 |
| T7 terminator | 3530 ~ 3577 |
| 6x His (His Tag) | 3446 ~ 3463 |
| Stop codon | 3464 |
| Amp ^r gene | 599 ~ 1459 |
| Ori | 1630 ~ 2218 |
| f1 ori | 12 ~ 467 |

DNA sequence of multiple cloning sites in T&A Expression Vector (C-terminal)

```

3301 CCCGTGGCCA GGACCCAACG CTGCCCGAGA TCTCGATCCC GCGAAATTAA
GGGCACCGGT CCTGGGTTGC GACGGGCTCT AGAGCTAGGG CGCTTAAATT
3351 TACGACTCAC TATAGGGGAGA CCACAACGGT TTCCCTCTAG AAATAATTTT
ATGCTGAGTG ATATCCCTCT GGTGTTGCCA AAGGGAGATC TTTATTAATA
3401 GTTAACTTT AAGAAGGAGA TATACCCATG AAT Insert DNA ACCGCTG
CAAATTGAAA TTCTTCCTCT ATATGGGTAC TTAMet Asn TGGCGAC
ThrAlaG
3441 GCGAGCACCA CCACCACCAC CACTGAGATC CGGCTGCTAA CAAAGCCCGA
CGCTCGTGGT GGTGGTGGTG GTGACTCTAG GCCGACGATT GTTTCGGGCT
lyGluHisHis sHisHisHis HisEnd
3491 AAGGAAGCTG AGTTGGCTGC TGCCACCGCT GAGCAATAAC TAGCATAACC
TTCTTCGAC TCAACCGACG ACGGTGGCGA CTCGTTATTG ATCGTATTGG
T7 terminator
3541 CCTTGGGGCC TCTAAACGGG TCTTGAGGGG TTTTTTGCTG AAAGGAGGAA
GGAACCCCGG AGATTTGCCG AGAACTCCCC AAAAAACGAC TTTCTCTCTT

```

Restriction enzymes that ONE Cut T&A Expression Vector (C-terminal)

| Enzyme | Position | Enzyme | Position | Enzyme | Position | Enzyme | Position | Enzyme | Position |
|--------|----------|--------|----------|---------|----------|----------|----------|---------|----------|
| AccI | 2511 | Bbr7I | 3155 | BseYI | 1976 | HaeIV | 1392 | PstI | 1146 |
| AfeI | 3028 | BbsI | 3155 | BsgI | 3105 | MscI | 3311 | PvuI | 1020 |
| AflIII | 2280 | BglI | 1273 | BsmBI | 2633 | NaeI | 143 | PvuII | 2691 |
| AhdI | 1392 | BglII | 3334 | BstZ17I | 2511 | NgoMIV | 143 | SapI | 2403 |
| Alol | 302 | BlnI | 3525 | BtgI | 3308 | Nli3877I | 3330 | Scal | 909 |
| AlwNI | 1869 | Bpu10I | 3175 | DrallI | 249 | PciI | 2280 | StyI | 3547 |
| AvaI | 3330 | BsaHI | 851 | FspAI | 3302 | PfoI | 2638 | Tth111I | 2538 |
| BanII | 173 | BsaXI | 300 | GdiII | 999 | PsiI | 374 | XbaI | 3392 |

YB Rapid Ligation Kit



FYC003-100R (100 rxns)

| | |
|-----------------------|-----------------|
| yT4 DNA Ligase | 100 µl (3 U/µl) |
| 10× Ligation Buffer A | 200 µl |
| 10× Ligation Buffer B | 200 µl |

Storage Buffer

| | |
|----------------------|--------|
| Tris-HCl (pH7.5) | 20 mM |
| KCl | 50 mM |
| DTT | 1 mM |
| EDTA | 0.1 mM |
| Glycerol stabilizers | 50% |

10× Ligation Buffer A

| | |
|---------------------|--------|
| Tris-HCl | 0.4 mM |
| MgCl ₂ | 0.1 mM |
| DTT | 0.1 mM |
| ATP (pH5.0 at 25°C) | 5 mM |

The performance of this buffer depends on the integrity of ATP. Store the buffer in small aliquots at -20°C to minimize degradation of ATP and DTT

10× Ligation Buffer B

Buffer contains an enhancer which dramatically increases ligation efficiency for blunt end DNA

Description

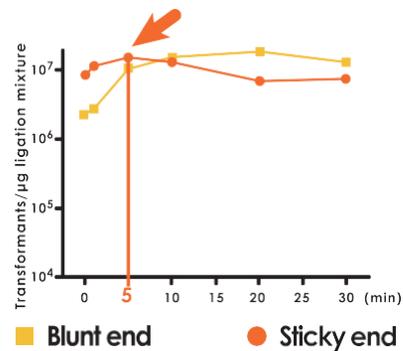
Yeastern's yT4 DNA ligase catalyzes the joining of two strands of DNA between the 5' - phosphate and the 3' - hydroxyl groups of adjacent nucleotides in either a cohesive-ended or blunt-ended termini. The enzyme also repairs single-strand nicks in duplex DNA, RNA or DNA/RNA hybrids. YB Rapid Ligation Kit is designed for efficient ligation of DNA inserts into vectors in just 5 minutes.

Features

- Either sticky- or blunt-end, ligation can be achieved within 5 minutes
- Reaction mixture can be used directly for bacterial transformation

Applications

- Join double-strand DNA with cohesive or blunt termini
- Join oligonucleotide linkers to DNA sequence with blunt end
- Repair nicks in duplex DNA, RNA, or DNA-RNA hybrids



Combo Products



Quality Control

Undetectable endo-deoxyribonuclease and exo-deoxyribonuclease activities, undetectable ribonuclease activity, undetectable degradation of labeled oligonucleotide, passed the test of the capability to join cohesive- and blunt-ended DNA fragments.

SCOS Transformation Kit

Description

The SCOS (*Saccharomyces cerevisiae* one-step) Transformation Kit provides a simple and fast one-step / one-tube method for transforming the yeast, *S. cerevisiae* with a linear or circular plasmid DNA. The transformation efficiency varies based on the yeast strain used, the efficiency of plasmid incorporated into the host chromosome, the selection marker used and the transformation procedure. In general, the entire procedure can be completed in one hour, and routinely provides a transformation efficiency of $10^3 - 10^6$ transformants/mg of plasmid DNA.

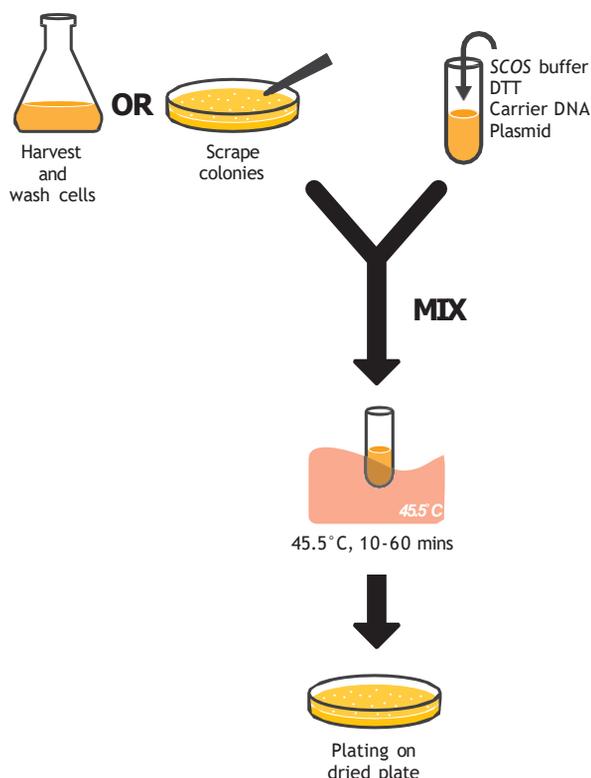
Features

- High throughput transformation
- Instant competent cell preparation
- An one-step, one-tube and 10-60 minutes protocol for transforming the baker's yeast (*Saccharomyces cerevisiae*)
- Simple (Mix → Heat shock → Plating)
- Suitable for yeast cells from colonies, broth or any growth phase
- Repeatable efficiency, always reach efficiency of $10^3 - 10^6$ transformants/ μg DNA

Applications

- Transformation of *S. cerevisiae*

Protocol



4°C

FYY101-120P (120 preps)

| | |
|----------------------------|---------|
| SCOS Transformation Buffer | 12 ml |
| Carrier DNA | 0.6 ml |
| DTT Powder | 0.185 g |

Quality Control

The transformation efficiency is checked in a lot-to-lot basis to ensure that it is above minimum 10^3 transformants/ μg of plasmid DNA.

YLEX Yeast Expression Kit



4°C

FYY201-1KT

| | |
|---|--------|
| <i>Yarrowia lipolytica</i> Yeast Strain: Po1g (# FYY202-1SB) | 1 stab |
| pYLEX1 - Expression Vector (# FYY203-5UG) | 5 µg |
| pYLSC1 - Secretion Vector (# FYY204-5UG) | 5 µg |
| Primer 6560 F (# FYY205-500UL) | 250 µl |
| Primer 6904 R | 250 µl |
| YLOS Yeast Transformation Kit (# FYY301-120P) | 1 kit |

Description

YLEX Yeast Expression Kit based on INRA INAPG licensed patent* provides an easy approach for cloning and expressing a gene of interest in the yeast, *Yarrowia lipolytica*. Using this kit, high level of heterologous protein may be expressed intracellularly or be secreted from the cell into medium by selecting the supplied expression vector pYLEX1 or pYLSC1.

Vectors provided in the YLEX contain a strong hybrid promoter carrying four tandem copies of upstream activator sequences (UAS1B) fragment from pXPR2 and a minimal pLEU2 fragment. Unlike the frequently used *Yarrowia* promoter (pXPR2), this stable hybrid promoter directs protein expression constantly without multiple influences by nutritional and environmental factors in medium.

When a constructed plasmid with the hybrid promoter followed by a cloned gene of interest is linearized by the selected restriction enzyme, it becomes an expression cassette that can integrate into the *Y. lipolytica* genome by homologous recombination within the process of transformation. The successful transformants are ready for expression or secretion of recombinant protein depending on whether secretion signal appears on the plasmid.

For more information, please read the articles cited in this catalog.

* INRA (Institut National de la Recherche Agronomique) and INAPG (Institut National Agronomique Paris-Grignon)

Features

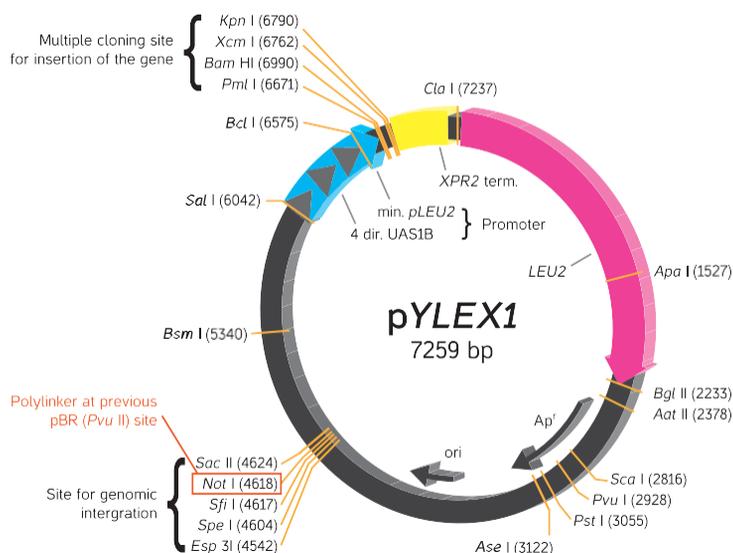
- **Safe:** *Y. lipolytica* was classified as GRAS (Generally Regarded As Safe) by the US FDA (Food and Drug Administration)
- **Simple:** a simple tool for expressing heterologous protein
- **Easy manipulation:** like *E. coli* and *S. cerevisiae*
- **Stable:** strong stability in vectors and constructed plasmids
- **Reliable:** vectors integrated at the same site in genome
- **Flexible:** both expression and secretion vector provided (proteins may be expressed intracellularly or be secreted from the cell into medium)
- **High growth ability:** high secretion capacity & high product yield
- **Less protein degraded:** no extracellular protease synthesized by a special protease-deficient *Yarrowia* strain
- **Mass production:** industrial mass production of recombinant proteins
- **Less hyperglycosylation:** able to perform post-translational processing of complex proteins, unlike *S. cerevisiae*

Applications

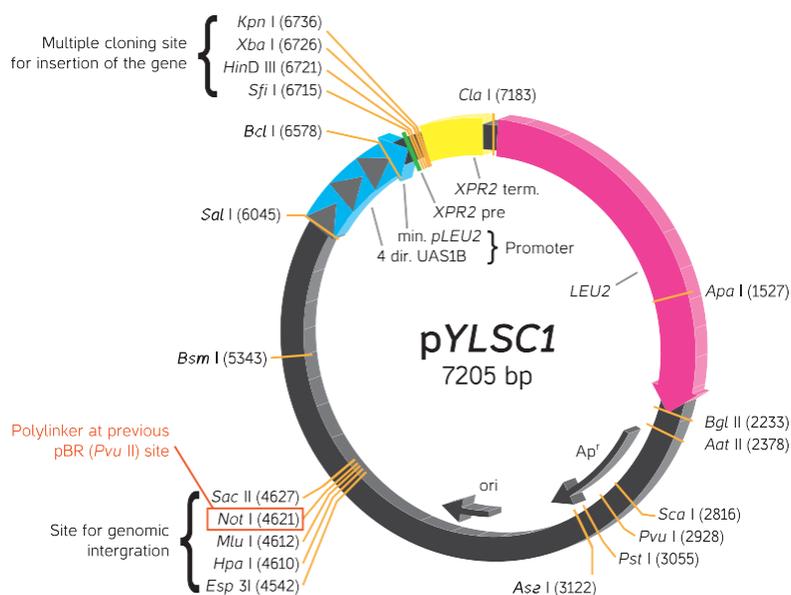
Heterologous protein expression, either intracellular or extracellular depends on selected vector in GRAS yeast.

Yarrowia Vectors

Two vectors (pYLEX1 and pYLSC1) are included in this kit, and they can be used for either intracellular expression or secretion of proteins of interest in *Y. lipolytica*. Generally speaking, if the target protein is cytosolic and non-glycosylated, the pYLEX1 vector is a better choice. If the protein of your interest is normally glycosylated or secreted, you may wish to choose the pYLSC1 vector.



The pYLEX1 expression vector (7259 bp) contains the strong hybrid promoter (hp4d) carrying four tandem copies of upstream activator sequences (UAS1B) fragment from pXPR2 and a minimal pLEU2 fragment. The multiple cloning site and the XPR2 transcription terminator lie immediately downstream of 3' site of hp4d promoter, followed by a leucine selection marker gene (LEU2). The vector can be linearized by digestion with *Not*I to create a linear DNA fragment capable of inserting into the *Y. lipolytica* genome.



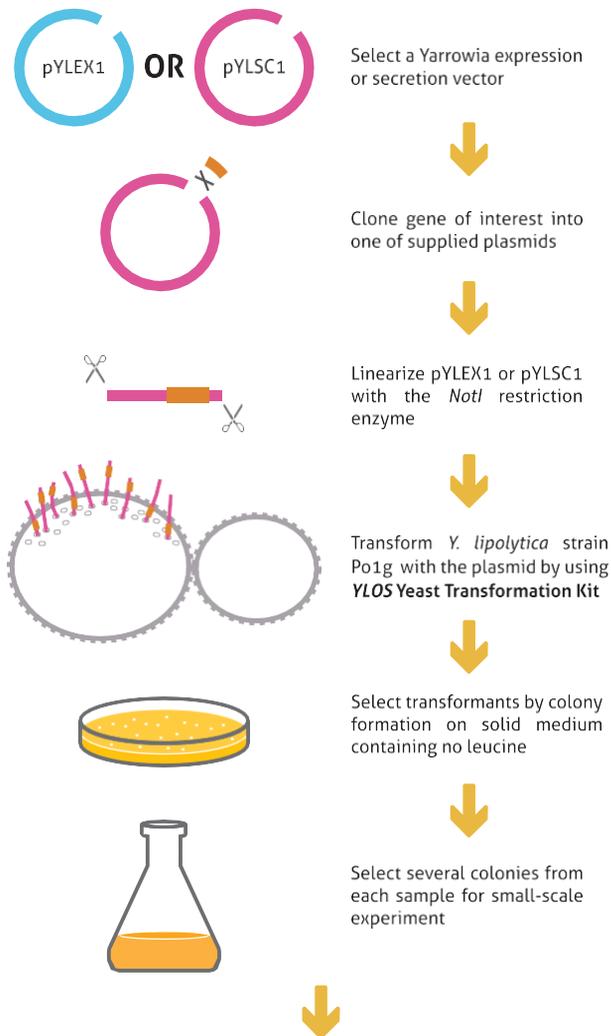
The pYLSC1 secretion vector (7205 bp) contains the hybrid promoter (hp4d) and a secretion signal (XPR2 pre region). The multiple cloning site and the pXPR2 transcription terminator lie immediately downstream of 3' site of XPR2 pre region, followed by a leucine selection marker gene (LEU2). The vector can be linearized by digestion with *Not*I to create a linear DNA fragment capable of inserting into the *Y. lipolytica* genome.

Yeast Strain

The strain Po1g of *Yarrowia lipolytica* is a derivative of the wild-type strain W29 (ATCC 20460) by a series of genetic modifications. Briefly, the original *URA1* gene in the W29 strain was disrupted with the *SUC2* gene from *Saccharomyces cerevisiae*, followed by the introduction of a deletion in the *LEU2* gene. Furthermore, the deletion of the *XPR2* and *AXP* genes ensures that Po1g is unable to produce any extracellular protease. In order to allow easy integration of pBR-based expression/secretion vectors, a pBR322 docking platform was integrated at the *URA1* locus.

| Strain | Genotype | Phenotype |
|--------|--|--------------------------------------|
| Po1g | <i>MatA</i> , <i>leu2-270</i> , <i>ura3-302::URA3</i> , <i>xpr2-332</i> , <i>axp-2</i> | Leu-, ΔAEP, ΔAXP, Suc+, pBR platform |

Protocol



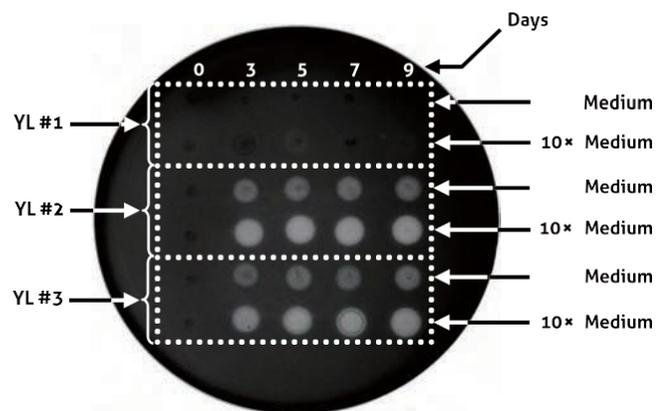
Analyze the protein of interest

Quality Control

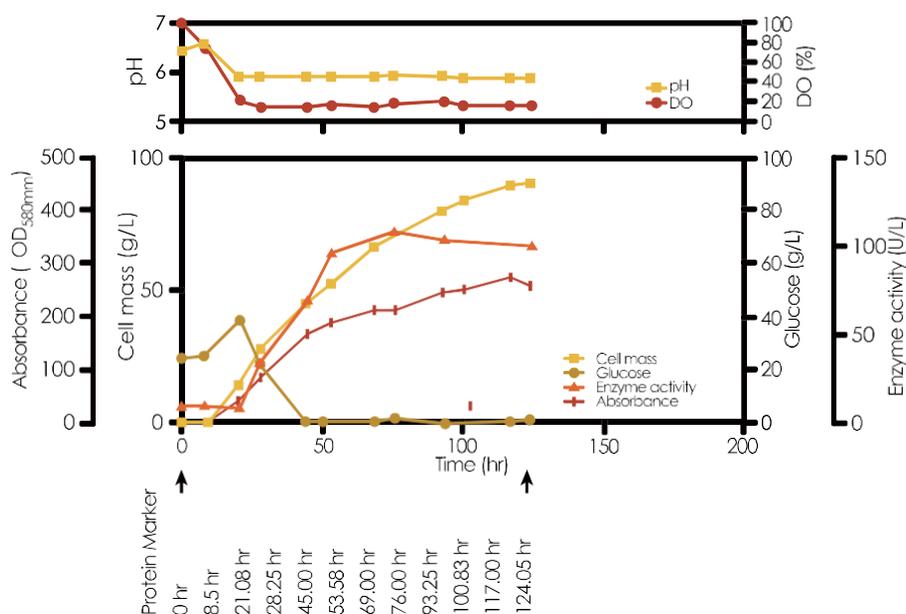
Two plasmids, pYLEX1+AMY1 (mouse salivary α -amylase gene) and pYLSC1+AMY1Δ (*AMY1* without its native secretion signal) are used to ensure functional transformation kit and correct gene expression in the host. Contamination test is also performed to ensure no other microbial contamination.

Experimental Data

The figure shows that filtered culture medium from batch culture of both amylase-encoding transformants (YL #2 and #3) could digest starch in solid medium agar, and subsequently produce clear zones. In contrast, medium from the culture of yeast transformed with vector only (YL #1) did not exhibit the same result. It indicates that cloning of both α -amylase gene (*AMY1*) into pYLEX1 and α -amylase gene without its secretion signal peptide (*AMY1*Δ) into pYLSC1 was successful by using the YLEX Yeast Expression Kit. In both cases, active α -amylase was efficiently secreted into the culture medium.

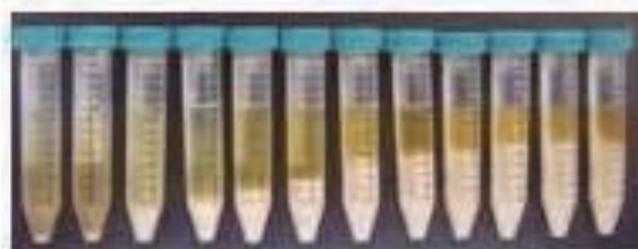


A time course study of capacity of high cell density fermentation and secretion of recombinant enzyme in the YLEX system (*Yarrowia lipolytica* expression system).



Over production of heterologous protein (SDS-PAGE analysis):

Secretion profile of recombinant enzyme in supernatant, samples were collected by time course, 10 µl/lane.



High cell density fermentation:

High cell density fermentation in a 14L fed-batch fermenter, cells were centrifuged (3,000 rpm) by time-course.

*** YLEX Yeast Expression Kit (product spec.)**

Yarrowia lipolytica yeast strain: Po1g; pYLEX1- expression vector 1 stab; pYLSC1- secretion vector 5 µg; Primer 6560F (for sequencing purpose) 250 µl; Primer 6904F (for sequencing purpose) 250 µl; YLOS Yeast Transformation Kit 1 kit (120 rxns).

* YLEX Yeast Expression Kit is produced by **Yeastern Biotech Co., Ltd.** with patent protected, which is authorized with the non-exclusive manufacturing and distribution worldwide by the proprietary property of Institute National de la Recherche Agronomique (INRA), France.

YLOS Yeast Transformation Kit



4°C

FYY301-120P (120 preps)

| | |
|----------------------------|---------|
| YLOS Transformation Buffer | 12 ml |
| Carrier DNA | 0.6 ml |
| DTT Powder | 0.185 g |

Description

The YLOS (*Yarrowia lipolytica* one-step) Yeast Transformation Kit provides a simple and fast one-step/one-tube method to transform *Yarrowia* cells cultured in either solid agar or liquid broth. They were designed for various strains of *Y. lipolytica* and vectors. Transformation efficiencies with *Y. lipolytica* will vary based on the yeast strain used, a linear or circular plasmid DNA, the efficiency of plasmid integration into the host chromosome, and the transformation procedure chosen. Generally, the entire procedure may be completed in 60 mins, and routinely provides a transformation efficiency of 10^3 - 10^5 transformants/ μ g of plasmid DNA.

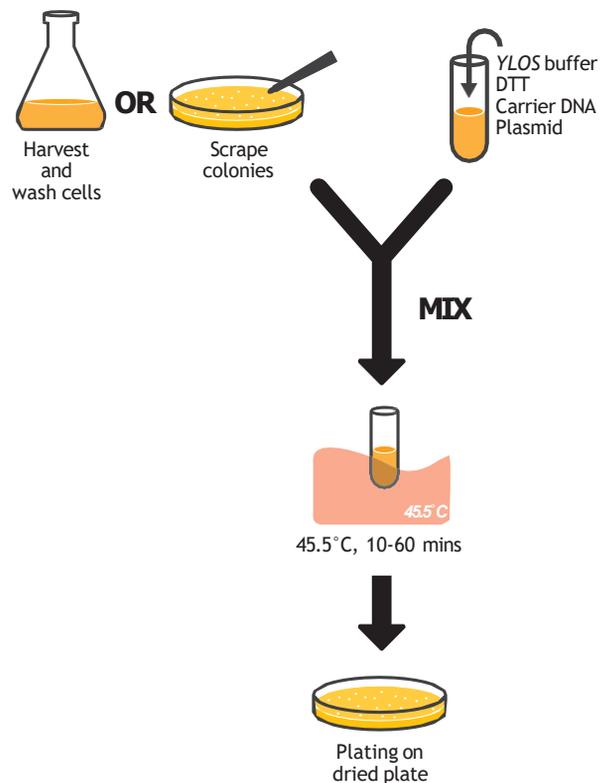
Features

- Instant preparation of competent cells
- An one-step, one-tube and 10 - 60 minutes protocol for transforming
- The oleaginous yeast, *Yarrowia lipolytica*
- Simple (Mix → Heat shock → Plating), suitable for *Yarrowia* yeast
- Cells from colonies, broth or any growth phase
- Repeatable efficiency, always reach efficiency of 10^3 - 10^5 transformants/ μ g DNA

Applications

- Transformation of *Y. lipolytica*

Protocol



Quality Control

The transformation efficiency with the plasmid pYLEX1 is checked in a lot-to-lot basis to ensure that it is above minimum 10^3 transformants/ μ g of plasmid DNA.

Glass Plating Beads (Sterilized)

Description

Sterilized glass plating beads (4 mm) provide an efficient and easy method to achieve optimal spreading on agar plates. The rolling action of the glass beads gently spread the *E. coli* solution (or any bacterial/fungal solution) to reach areas of the plate that are inaccessible to spread. Multiple plates containing beads and *E. coli* solution may be stacked and shaken simultaneously to increase spreading throughout and reduce cell stress. The beads may be recovered by washing with 70% ethanol, then sterilized and dried.



RT

FYO001-100G

Sterilized, 4 mm

100g

Features

- Pre-sterilized
- No flaming required

Applications

- Bacterial and fungal solution spreading on agar plates
- Plating yeast for two-hybrid screens

Protocol

For regular plating of ECOS™ Competent Cells, add 5 ~ 10 beads immediately after pipetting 50 ~ 100 µl of transformed cells onto each plate. Move the plate back and forth to roll the beads on the surface and change the direction of shaking a few times to ensure even spreading. When the surface of the agar starts to dry on the plate (20 ~ 40 seconds), pour off the beads and the plates are ready for incubation.

- Pre-warmed glass beads at 37°C for ECOS™ warm plating (protocol 2&4, page 3-2 protocol)
- Pre-cooled glass beads at 4°C for ECOS™ cold plating (protocol 1&3, page 3-2 protocol)
- Room temperature for SCOS and YLOS Yeast Transformation Kits

Quality Control

Each lot of beads has to pass contamination test by spreading sterilized water on agar plates containing no selective antibiotics. No bacterial and fungal colonies should be seen after 2 days incubation.

Related Products

- ECOS™ Competent cells

3-1

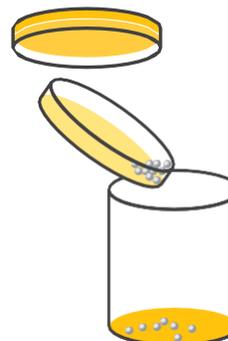
Protocol



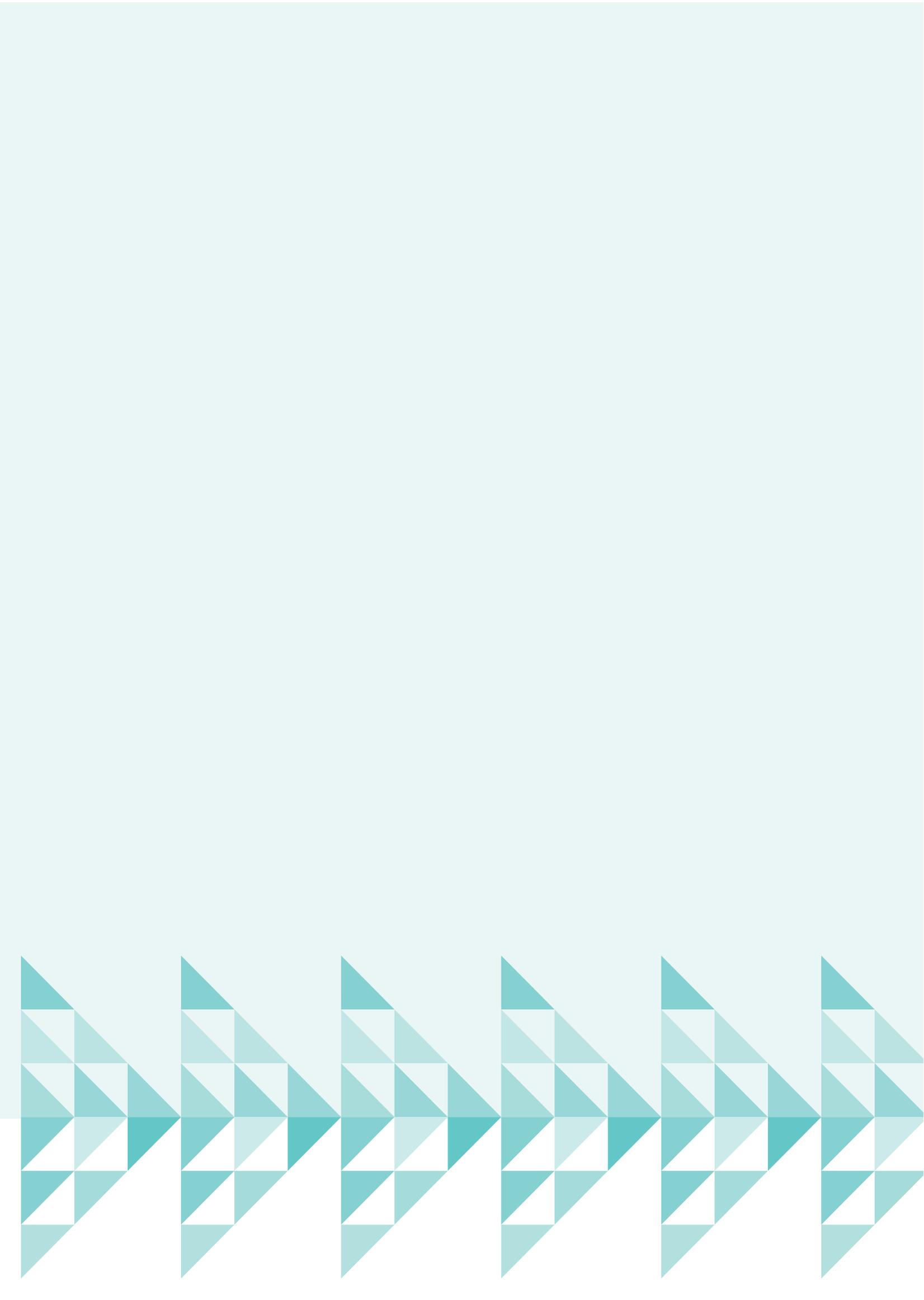
Add beads after pipetting transformed cells onto each plate.

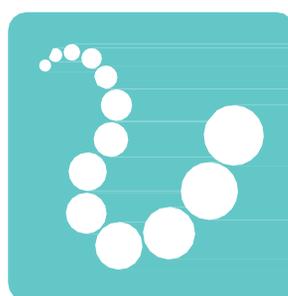


Shake the plate to roll the beads on the surface and change the direction of shaking a few times to ensure even spreading.



When the surface of the agar starts to dry on the plate (20 ~ 40 seconds), pour off the beads and the plates are ready for incubation.





Proteomics

SupraNase Nuclease



-20°C

FYP301-5KU

100 µl

FYP301-25KU

500 µl

FYP301-100KU

Powder

Description

SupraNase (EC: 3.1.30.2), is derived from the bacterium *Serratia Marcescens*. It cleaves both single and double stranded DNA and RNA, whether linear or circular. This recombinant endonuclease is expressed and purified from *E. coli* as a non-covalent homo-dimer. Each chain consists of 245 amino acids with a molecular weight of ~29 kD. It is primarily used to remove residual nucleic acid contaminants from biological expression systems and reduce viscosity in cell lysates during purification processes for proteins, vaccines, and antibodies.

Applications

- **Biopharmaceutical manufacturing:** Reduction of host-cell DNA/RNA during downstream processing of recombinant proteins, vaccines, and viral vectors.
- **Viscosity reduction:** Improves filtration, centrifugation, and chromatography by lowering lysate viscosity.
- **Research and laboratory use:** Removal of nucleic acid contamination during protein purification and sample preparation.
- **Industrial biotechnology:** Used in fermentation and cell-lysis workflows to enhance product recovery.

Operating Conditions

The recommended reaction conditions for cleavage by SupraNase are as follows: 50 mM Tris buffer with 1-2 mM Mg²⁺ at a pH of 8-9, and a temperature of 37 °C. However, SupraNase can tolerate high concentrations of reductants such as DTT and denaturants like urea and guanidine hydrochloride, while maintaining its high activity. This versatility allows SupraNase to be used in various sample conditions.

Preparation of nuclease solution:

- If powder form: To prepare the nuclease solution, add 1 ml deionized water and 1 ml 100% glycerin to each vial containing lyophilized powder, resulting in a solution ranging from 50 U/µL. Gently wash the vial along the wall using a pipette tip. If needed, the vial can be inverted several times with the cap covered, followed by centrifugation for later use or further dilution with a cleavage buffer.
- Sample processing for protein purification: Typically, 100-500 units of SupraNase are used per gram of cell paste. Add SupraNase before performing lysis. Incubate the mixture with gentle shaking or rotation for 10-20 minutes at room temperature. If necessary, the enzymatic reaction can be optimized.
- Sample processing for bio-analysis of proteins: For processing animal tissue samples in bio-analysis, sonication on ice is performed in the presence of SupraNase (50 U/100 µl) using a lysis buffer. Sonication is repeated every 15 minutes for 1 hour, followed by centrifugation at 750 g and 4°C for 5 minutes to remove debris.

Related Products

- ECOS™ Competent cells 3-1
- T&A™ Expression Vector 3-17

Note





OEM Services & Appendix

Competent Cells

Description

We are the world's leading competent cells manufacturing biotechnology company, providing customers from all over the world with reliable and quality competent cells. Yeastern Biotech Co., Ltd. owns the patent of ECOS™ technology (including protocol) exclusively in Canada (**TMA622,671**), USA (**US 6,864,088, US 7,098,033, US 7,820,443**), UK (**GB2383582**), German (**Nr. 102 51 429**), France (**FR 2832727**), Taiwan (**I 229696**), China (**ZL 2005 1 0112590.8**), Korea (**0604787, 10-1350283**), and Japan (**4867595**). Under the patented protocol, transformation with our ECOS™ chemically competent cells can be finished in one step within one minute without the need for SOC recovery step. Therefore, ECOS™ competent cells are the fastest transforming cells worldwide.

In order to meet customer needs, we provide OEM of *E. coli* competent cells using our ECOS™ technology. Any *E. coli* strain can be prepared to become ECOS™ competent cells which can be transformed using our patented protocols. Customers will only need to send the *E. coli* strain to us along with information of its genotype and note for basic culture conditions. Typically, the total procedure can be finished in 30-60 days. However, a minimum order of 160 tubes (100 µl/tube) is required for this service.

Molecular Biology Kits

Description

Yeastern have completed many OEM requests on the development of molecular biology kits, such as competent cells, DNA ladders, DNA polymerases, PCR premixes, RT-PCR premixes, qPCR premixes, pre-stained protein markers, fast DNA detection or diagnostic kits. We sincerely contribute our technologies through OEM services to satisfy customers' requirements.

Yeast Genetic Engineering

Saccharomyces cerevisiae

Yeastern's genetic engineering system with food-grade baker's yeast (*Saccharomyces cerevisiae*) mutants (patent pending) enables secretion or accumulation of high-quality heterologous proteins at either analytical or industrial scales. It is designed to lessen the risk and cost of pharmaceutical and biotechnology companies in high-quality protein production. Currently, we have completed the design and production of more than 20 proteins (peptide hormones, transmembrane proteins, industrial enzymes, nucleic acid binding proteins, drug proteins, viral antigens), and most have shown low glycosylation and high production. For our own, we have successfully produced herbal immunomodulatory proteins and algal SOD using this technology.

Yarrowia lipolytica

Yarrowia lipolytica is classified as GRAS (Generally Regarded As Safe) by the US FDA (Food and Drug Administration). *Y. lipolytica*, like *E. coli* and *S.cerevisiae*, can be manipulated easily and produces proteins with less hyperglycosylation. *Y. lipolytica* genetic engineering system is based on our YLEX Yeast Expression Kit, by which high level of heterologous proteins can be expressed intracellularly or be secreted from cells into the medium depending on customers' desires.

Educational Packs

Description

Yeastern manufactures a series of molecular biology products from nucleic acid extraction, amplification to gene cloning. We are able to OEM user-friendly educational packs, aiming to give high school and undergraduate students a fundamental knowledge of life science thus increase their interests in further exploring the fun and excitement of science!

Competent Cells Information

Transformation Efficiency

- Equation for transformation efficiency = transformed colonies (transformants) / μg of plasmid

- Example :**

100 μl of competent cells have been transformed with 10⁻⁶ μg of pUC19 plasmid. If 550 colonies are observed on the selective plate. The transformation efficiency is:

$$550 / 10^{-6} = 5.5 \times 10^8 \text{ transformants} / \mu\text{g of pUC19 plasmid}$$

Genotype

| Product Name | Strain | Genotype |
|----------------------------|-------------------|---|
| ECOS™ X / ECOS™ 101 | DH5 α | F ⁻ endA1 hsdR17(rk ⁻ , mk ⁻) supE44 thi-1 λ ⁻ recA1 gyrA96 relA1 Δ (argF-lacZYA)U169 Φ 80d lacZ Δ M15 deoR |
| ECOS™ 9-5 | JM109 | F' traD/6 proA ⁺ proB ⁺ lacIq Δ (lacZ)M15 Δ (lac-proAB) supE44 hsdR17 recA1 gyrA96 thi ⁻ 1 endA1 relA1 e14 ⁻ λ ⁻ |
| ECOS™ Blue | XL1-Blue | F' recA1 endA1 gyrA96 thi ⁻ 1 hsdR17(rk ⁻ , mk ⁺) supE44 λ ⁻ Δ (lac) proAB lacIq Δ M15 Tn10 (tet ^r) |
| ECOS™ 21 | BL21(DE3) | F ⁻ hsdS gal (λ clts857 ind1 Sam7 nin5 lacUV5-T7 gene l) |
| ECOS™ 10B | DH10B | F ⁻ endA1 recA1 galE15 galK16 nupG rpsL Δ lacX74 Φ 80d lacZ Δ M15 araD1/9 Δ (lara,leu)7697 mcrA Δ (mrr ⁻ hsdRMS ⁻ mcrBC) λ ⁻ |
| ECOS™ 2163 | GM2163 | F ⁻ ara ⁻ 14 leuB6 thi ⁻ 1 fhuA/1 lacY1 tsx ⁻ 78 galK2 galT22 supE44 rpsL1/6(strr) xyl ⁻ 5 mtl ⁻ 1 dam11:Tn9 (camr) dcm ⁻ 6 mcrB1 hsdR2(rk ⁻ mk ⁺) mcrA |
| ECOS™ St | HB101 Derived | F ⁻ mcrB mrr hsdS20(rB ⁻ , mB ⁻) recA13 supE44 ara-14 galK2 lacY1 proA2 rpsL20(Str ^R) xyl-5 λ ⁻ leu mtl-1 |
| ECOS™ SONIC | BL21(DE3) Derived | F ⁻ ompT hsdS _B (r _B ⁻ m _B ⁻) gal dcm (DE3) Δ endA Δ recA pLysS (Cam ^R) |

Nomenclature & Abbreviations

| Gene | Description |
|-------------|---|
| araD | Arabinose metabolism is blocked due to mutation in L-ribulose-phosphate 4-epimerase. |
| dam | Elimination of endogenous adenine methylation at GATC sequences. |
| dcm | Elimination of endogenous cytosine methylation at CCWGG sequences. |
| DE3 | A λ prophage carrying the T7 RNA polymerase gene and lacI ^q . |
| deoR | A regulatory gene that allows constitutive expression of deoxyribose synthesis genes; permits uptake of large plasmids. |
| endA | Eliminate the nonspecific activities digestion of Endonuclease I to obtain a cleaner preparation of DNA. |
| e14 | Excisable prophage-like element, containing mcrA gene, present in K-12 but missing from many derivatives. |
| F | The F plasmid, a low-copy number and self-transmissible plasmid. |

| Gene | Description |
|-------------------------|--|
| <i>fhuA</i> | Resistance to phage T1 (ferric hydroxamate uptake) due to mutation in iron uptake receptor. |
| <i>gal</i> | The ability to metabolize galactose is eliminated. |
| <i>galE</i> | A mutation which prevents the production of UDP-galactose, increased resistance to phage P1 infection and 2-deoxygalactose. <i>galE15</i> is a point mutation resulting in a Ser123 → Phe conversion. |
| <i>galK</i> | A mutation which prevents the metabolization of galactose and are resistant to 2-deoxygalactose. <i>galK16</i> is an IS2 insertion downstream of the <i>galK</i> start codon. |
| <i>gyrA</i> | Resistance to the antibiotic nalidixic acid due to a point mutation in DNA gyrase, subunit A. |
| <i>hsdR, hsdS</i> | DNA without methylation of certain sequences will be recognized as foreign and degraded by EcoK I or EcoB I. <i>hsdR</i> and <i>hsdS</i> recognize different sequences and are encoded by different alleles of <i>hsdRMS</i> . <i>hsdR</i> mutations eliminate restriction but not protective methylation (r-m+), while <i>hsdS</i> mutations eliminate both (r-m-). |
| <i>lacI^r</i> | Mutation of -35 site in upstream promoter of <i>lacI</i> , causing overproduction of <i>lac</i> repressor, result in turning off expression from Plac more completely. |
| <i>lacZ</i> | β-galactosidase activity is eliminated. |
| <i>lacZΔM15</i> | Deletion mutation of the <i>lacZ</i> gene (a.a. 11-41) that allows α complementation of the β-galactosidase gene; required for blue/white selection on XGal plates. |
| <i>lacY</i> | Lactose permease activity eliminated. |
| <i>lon</i> | Elimination of a protease activities responsible for degrading aberrant proteins. |
| <i>mcrA</i> | Mutation eliminating restriction of DNA methylated at the sequence C ^m CGG. |
| <i>mtl</i> | The ability to metabolize the sugar alcohol mannitol is eliminated. |
| <i>ompT</i> | Activity of outer membrane protease VII is eliminated, reducing proteolysis of expressed protein. |
| <i>phoA</i> | Activity of alkaline phosphatase is eliminated. |
| <i>recA</i> | Reduce unwanted homologous recombination. |
| <i>relA</i> | Permits RNA synthesis in the absence of protein synthesis. |
| <i>rfbD</i> | Unable to synthesis cell surface O-antigen as lack of functional TDP-rhamnose synthetase. |
| <i>rpsL</i> | Mutation in ribosomal protein S12 conveying streptomycin resistance. |
| <i>supE</i> | A tRNA functioning in suppressing the amber (UAG) stop codon by insertion of glutamine; required for growth of some phage vectors. |
| <i>thi-1</i> | The ability to synthesize thiamine (vit. B1) is eliminated. |
| <i>Tn10</i> | Transposon normally carrying Tetracycline resistance. |
| <i>tsx</i> | Resistance to bacteriophage T6 and colicin K. |
| (Φ80) | Strain carries the lambdoid prophage Φ80. A defective Φ80 prophage carrying the <i>lacM15</i> deletion is present in some strains. |

Nucleic Acids & Amino Acids Information

Concentration of Nucleic Acids

- **dsDNA (double-stranded DNA)** : 1 OD260 Unit = 50 µg/ml
- **ssDNA (single-stranded DNA)** : 1 OD260 Unit = 37 µg/ml
- **ssRNA (single-stranded RNA)** : 1 OD260 Unit = 40 µg/ml
- **dsDNA (double-stranded DNA)** : 1 OD260 Unit = 50 µg/ml
- **single-stranded oligonucleotides** : 1 OD260 Unit = 20 µg/ml

Molecular Weight Conversion of Nucleic Acids

Exact M.W. of ssRNA :

$$\text{M.W.} = (A_x \times 329.2) + (U_x \times 306.2) + (C_x \times 305.2) + (G_x \times 345.2) + 159^*$$

X : the number of each respective nucleotide within the polynucleotide.

* :M.W. of a 5' triphosphate.

Exact M.W. of ssDNA :

$$\text{M.W.} = (A_x \times 313.2) + (T_x \times 304.2) + (C_x \times 289.2) + (G_x \times 329.2) + 79.0^*$$

X : the number of each respective nucleotide within the polynucleotide.

* :M.W. of the 5' monophosphate left by most restriction enzymes. No phosphate is present at the 5' end of strands made by primer extension.

Molecular Weight of Amino Acids

| Amino Acid | 3-letter Code | 1-letter Code | Molecular Weight (g/mol) |
|------------|---------------|---------------|--------------------------|
| Alanine | Ala | A | 89.1 |
| Arginine | Arg | R | 174.2 |
| Asparagine | Asn | N | 132.1 |
| Aspartate | Asp | D | 133.1 |
| Cysteine | Cys | C | 121.2 |
| Glutamate | Glu | E | 147.1 |
| Glutamine | Gln | Q | 146.2 |
| Glycine | Gly | G | 75.1 |
| Histidine | His | H | 155.2 |
| Isoleucine | Ile | I | 131.2 |

| Amino Acid | 3-letter Code | 1-letter Code | Molecular Weight (g/mol) |
|---------------|---------------|---------------|--------------------------|
| Leucine | Leu | L | 131.2 |
| Lysine | Lys | K | 146.2 |
| Methionine | Met | M | 149.2 |
| Phenylalanine | Phe | F | 165.2 |
| Proline | Pro | P | 115.1 |
| Serine | Ser | S | 105.1 |
| Threonine | Thr | T | 119.1 |
| Tryptophan | Trp | W | 204.2 |
| Tyrosine | Tyr | Y | 181.2 |
| Valine | Val | V | 117.1 |

Gel Information

Agarose Gel

| %Agarose | Effective Range of Separation (bp) |
|----------|------------------------------------|
| 0.5 | 1,000 to 30,000 |
| 0.7 | 800 to 12,000 |
| 1.0 | 500 to 10,000 |
| 1.2 | 400 to 7,000 |
| 1.5 | 200 to 3,000 |
| 3-4 | 10 to 1000 |

Polyacrylamide Gel (Denaturing Gel)

| %Acrylamide | Effective Range of Separation (nucleotides) |
|-------------|---|
| 3.5 | > 500 |
| 5 | 151-500 |
| 10 | 61-150 |
| 15 | 30-60 |
| 20 | < 30 |

Note

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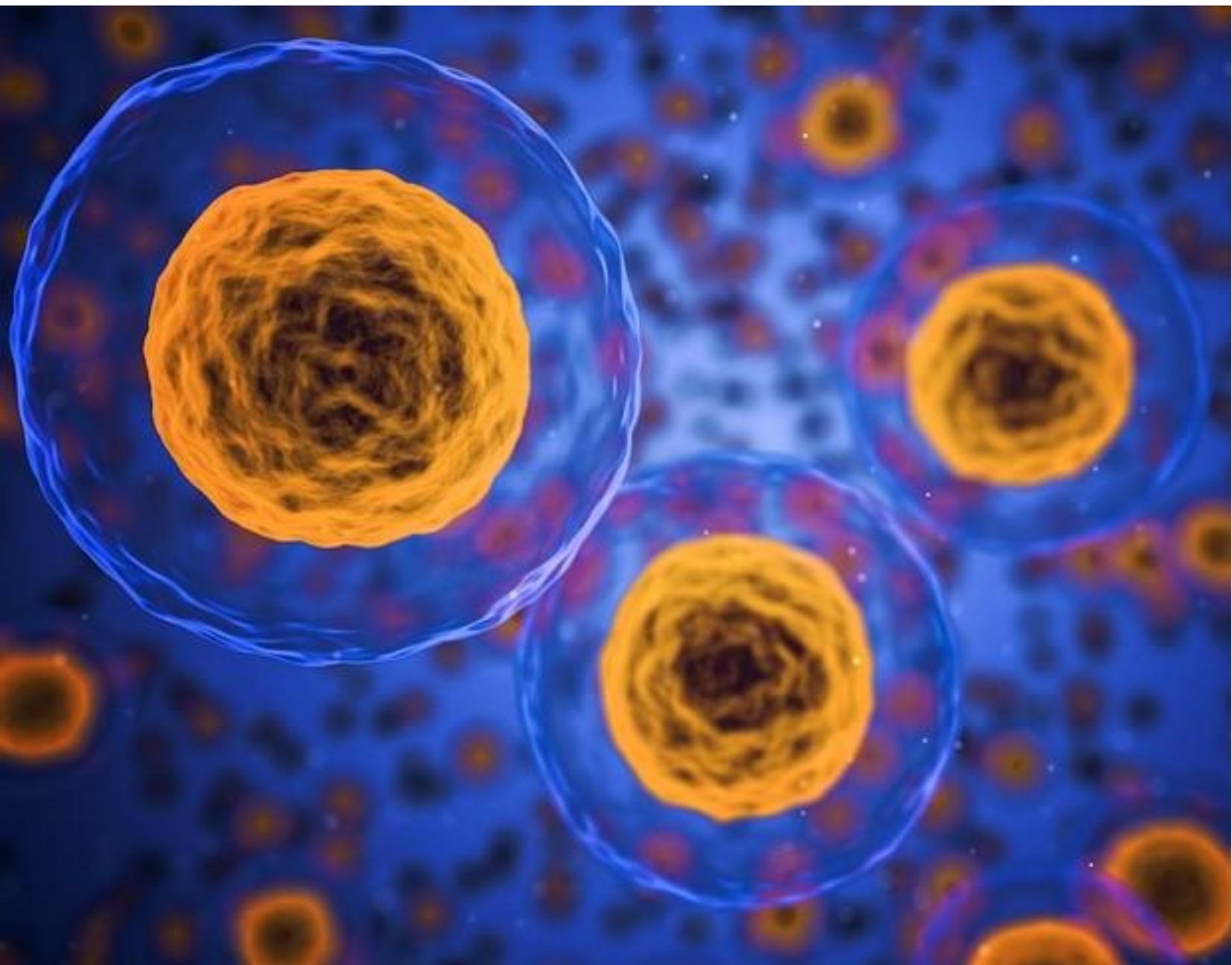
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